

# **Probabilistic environmental risk assessment of five nanomaterials (nano-TiO<sub>2</sub>, nano-Ag, nano-ZnO, CNT, and Fullerenes)**

Claudia Coll<sup>1</sup>, Dominic Notter<sup>1</sup>, Fadri Gottschalk<sup>2</sup>, Tianyin Sun<sup>1</sup>, Claudia Som<sup>1</sup>, Bernd Nowack<sup>1\*</sup>

1) EMPA - Swiss Federal Laboratories for Material Science and Technology, Technology and Society Laboratory, Lerchenfeldstrasse 5, CH-9014 St. Gallen, Switzerland.

2) ETSS – Environmental, technical and scientific services, Chaflur 136B, CH- 7558 Strada, Switzerland

*\*Corresponding author:*

*E-mail address: [nowack@empa.ch](mailto:nowack@empa.ch)*

*Tel.: +41 (0)58 765 76 92*

## **Supporting Information**

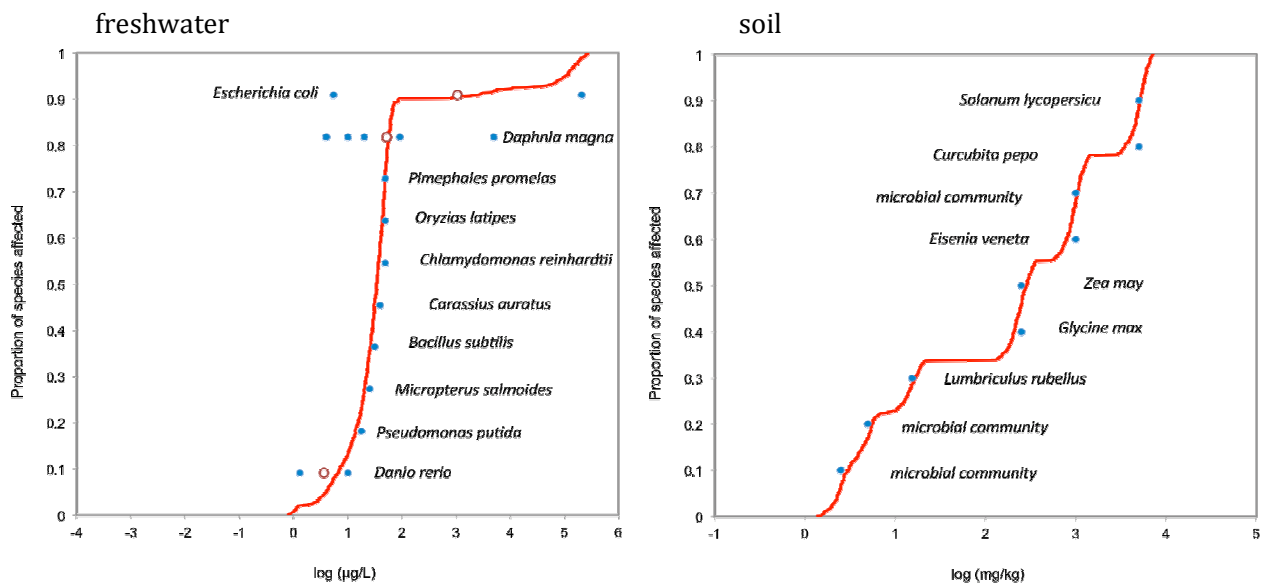


Figure S1: Probabilistic Species Sensitivity Distributions (PSSD) for fullerenes in the freshwater and soil compartments.

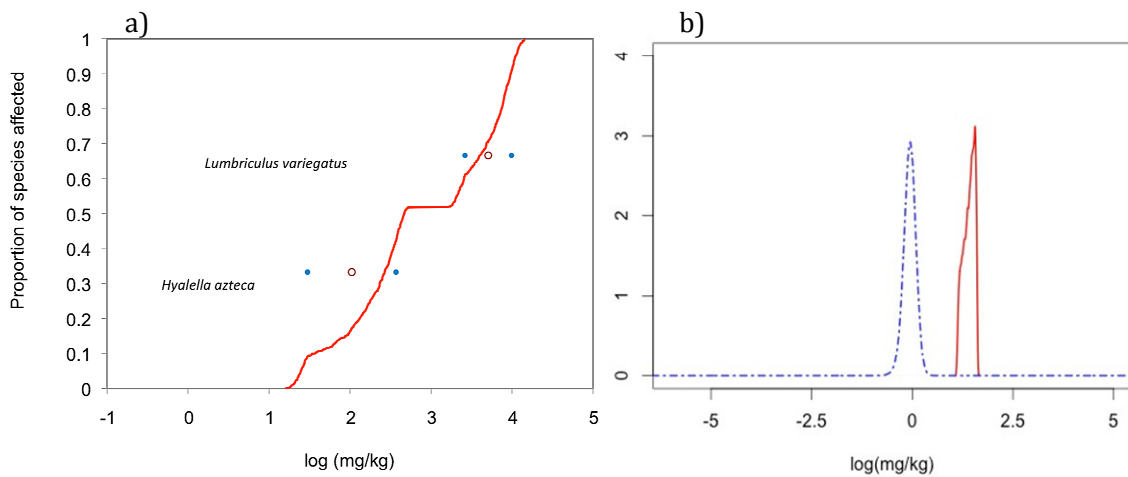


Figure S2: a) Probabilistic Species Sensitivity Distributions (PSSD) for CNTs in the sediment compartment. b) Comparison of predicted environmental concentrations (PEC) in blue and probabilistic species sensitivity distributions (pSSD) in red.

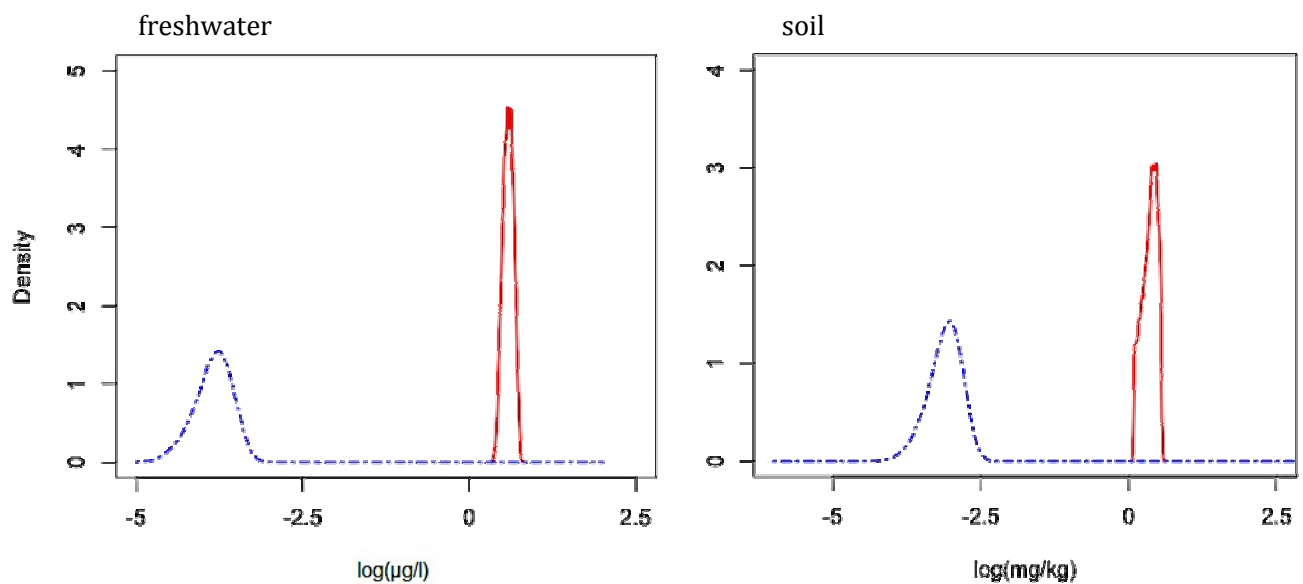


Figure S3: Comparison of predicted environmental concentration (PEC) in blue and probabilistic species sensitivity distribution (pSSD) in red for freshwater and sludge treated soils for fullerenes.

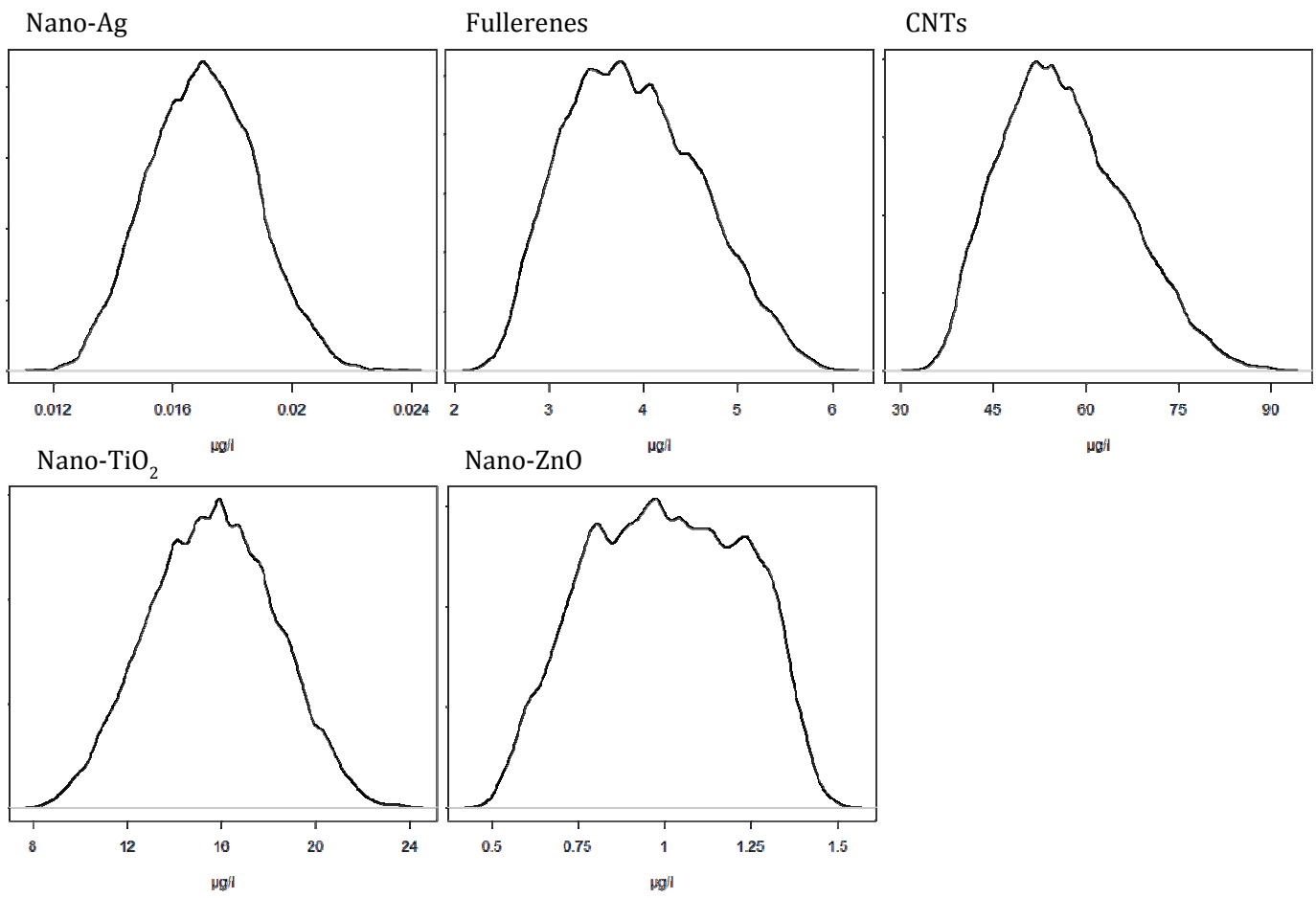


Figure S4: Predicted no-effect concentration distributions for TiO<sub>2</sub>, Ag, ZnO, fullerenes and CNT in freshwater.

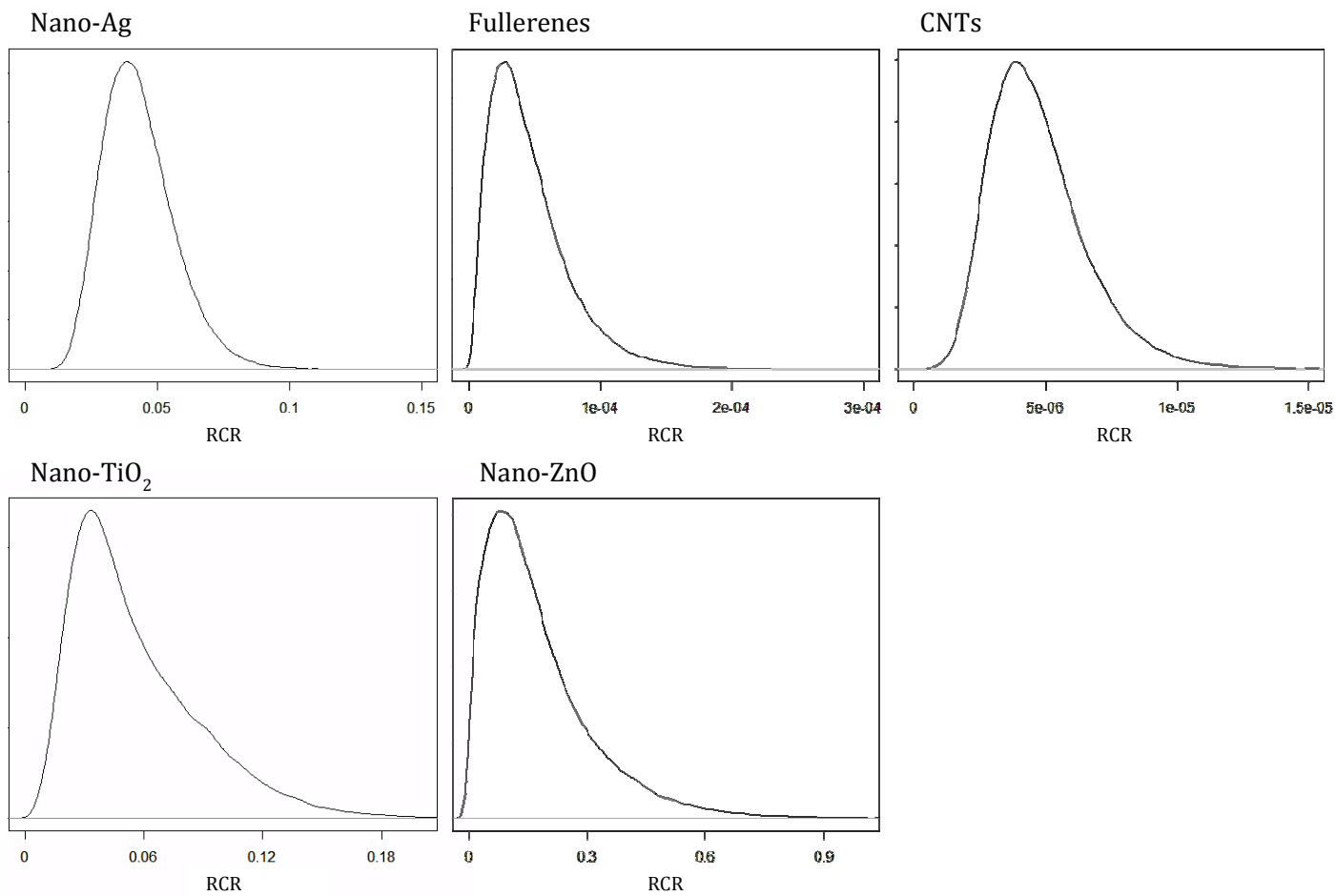


Figure S5: Risk Characterization Factor distributions for TiO<sub>2</sub>, Ag, ZnO, fullerenes and CNT in freshwater.

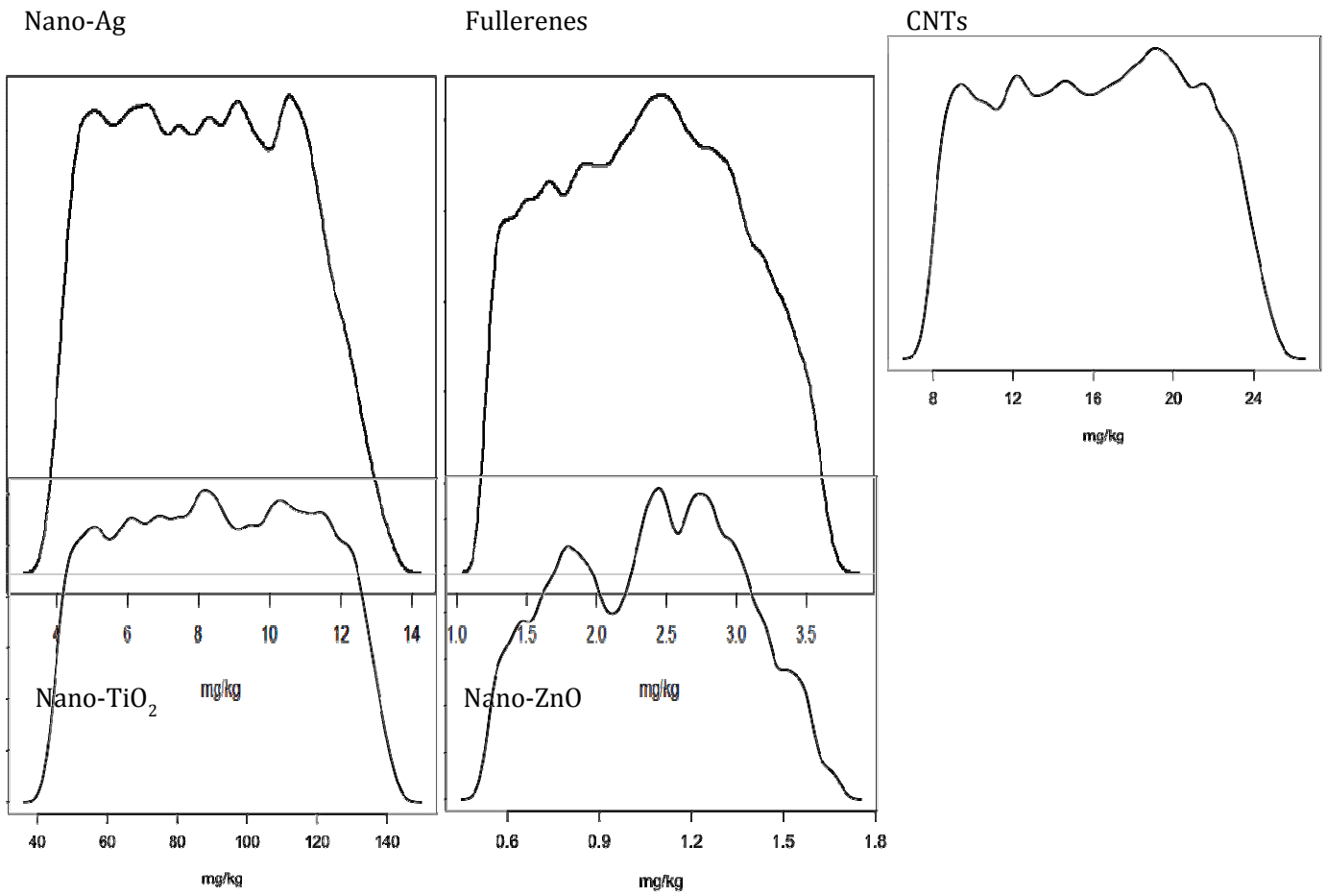


Figure S6: Predicted no-effect concentration distribution for TiO<sub>2</sub>, Ag, ZnO, fullerenes and CNT in soil.

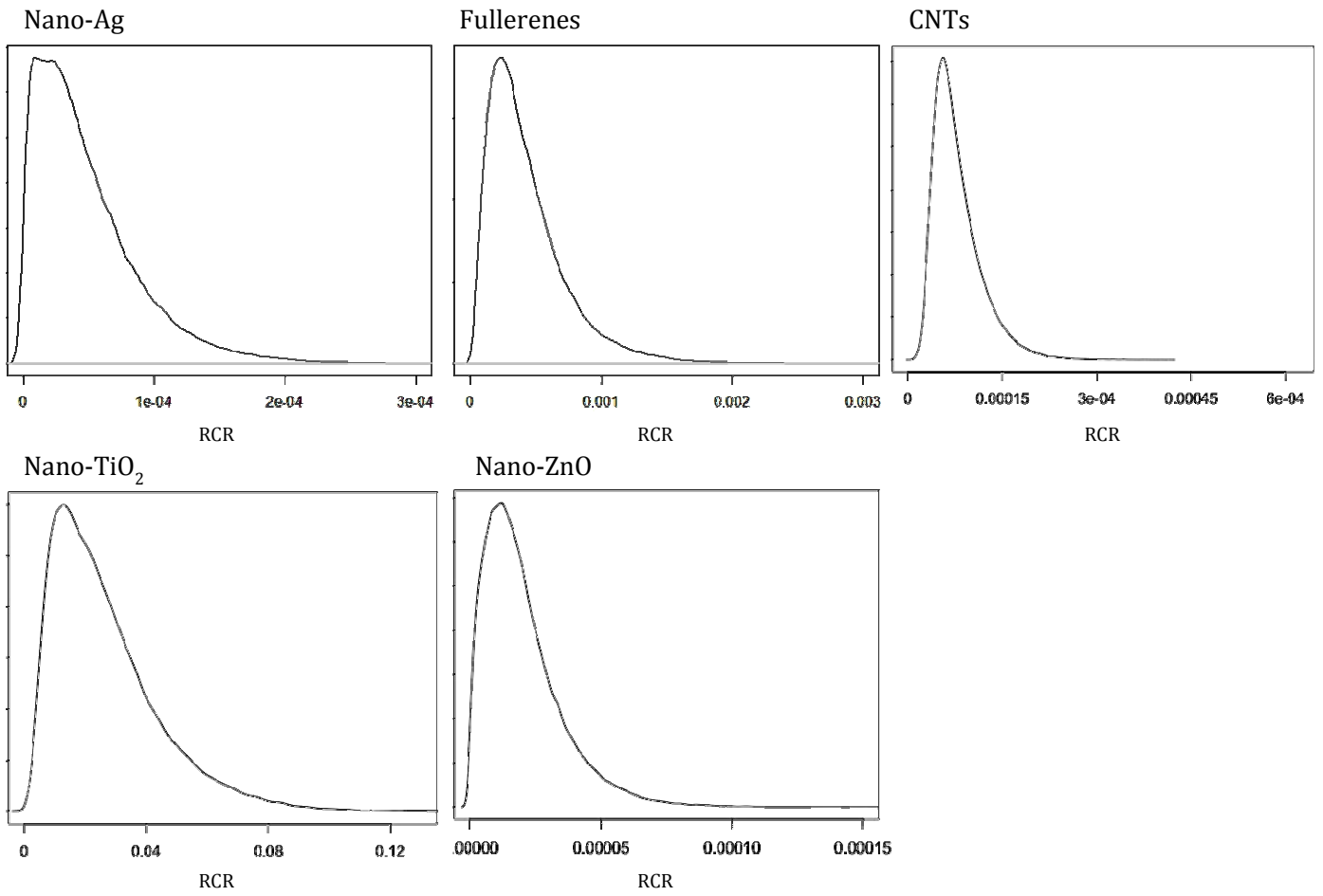


Figure S7: Risk Characterization Factor distributions for TiO<sub>2</sub>, Ag, ZnO, fullerenes and CNT in soil.

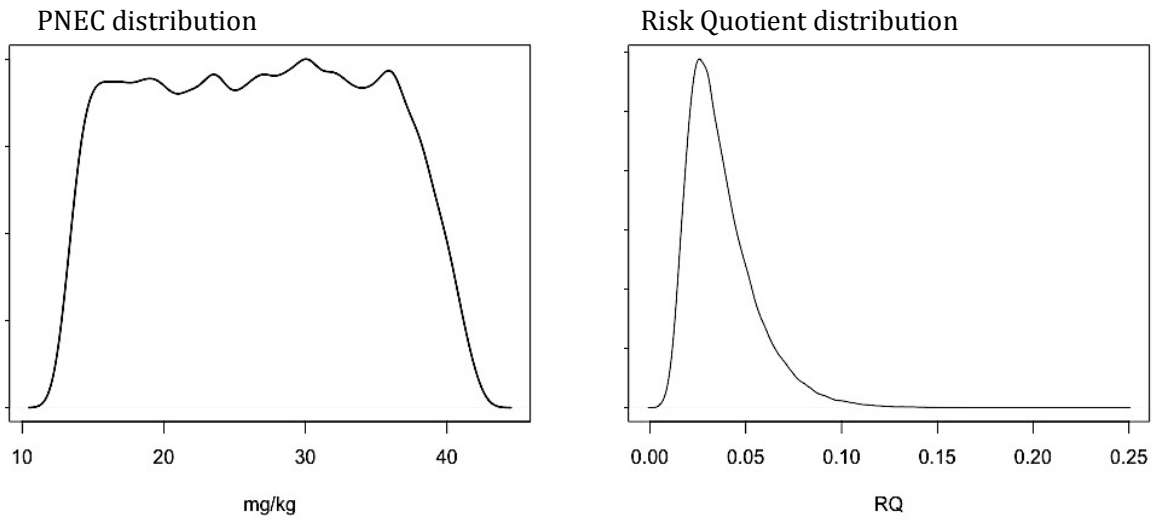


Figure S8: Predicted no-effect concentration distribution and risk quotient distribution for CNTs in sediment.



Table S1: Data for aquatic toxicity for nano-Ag, nano-TiO<sub>2</sub>, nano-ZnO, CNT and Fullerenes.

Author	ENM	Dose descriptor	Test organism	Concentration (µg/l)	Exposure time (h)	AF time	AF no-effect	Species sensitivity (µg/l)
Mouchet, et.al. [1]	CNT	HONEC	<i>Ambystoma mexicanum</i>	100,000.00	288	10	1	10,000.00
Templeton et al. [2]	CNT	NOEC	<i>Amphiascus tenuiremis</i>	1,600.00	720	1	1	1,600.00
Li et al. [3]	CNT	LC50	<i>Ceriodaphnia dubia, crustacea</i>	8,000.00	24	10	10	80.00
Alloy, et.al. [4]	CNT	LOEC	<i>Ceriodaphnia dubia, crustacea</i>	2,380.00	168	10	2	119.00
Kennedy et al. [5]	CNT	EC50	<i>Ceriodaphnia dubia, crustacea</i>	50,900.00	48	10	10	509.00
Long [6]	CNT	IC50	<i>Chlorella sp.</i>	12,400.00	96	1	10	1,240.00
Schwab et al. [7]	CNT	NOEC	<i>Chlorella vulgaris</i>	42.00	96	1	1	42.00
Schwab et al. [7]	CNT	NOEC	<i>Chlorella vulgaris</i>	180.00	96	1	1	180.00
Schwab et al. [7]	CNT	NOEC	<i>Chlorella vulgaris</i>	1,000.00	96	1	1	1,000.00
Schwab et al. [7]	CNT	NOEC	<i>Chlorella vulgaris</i>	3,000.00	96	1	1	3,000.00
Schwab et al. [7]	CNT	NOEC	<i>Chlorella vulgaris</i>	3,000.00	96	1	1	3,000.00
Asharani et al. [8]	CNT	NOEC	<i>Danio rerio, Zebrafish</i>	40,000.00	72	10	1	4,000.00
Cheng et al. [9]	CNT	LOEC	<i>Danio rerio, Zebrafish</i>	120,000.00	72	10	2	6,000.00
Zhu et al. [10]	CNT	EC50	<i>Daphnia magna, crustacea</i>	1,306.00	48	10	10	13.06
Edgington et al. [11]	CNT	LC50	<i>Daphnia magna, crustacea</i>	2,000.00	96	10	10	20.00
Zhu et al. [10]	CNT	EC50	<i>Daphnia magna, crustacea</i>	8,726.00	48	10	10	87.26
Alloy, et.al. [4]	CNT	LOEC	<i>Daphnia magna, crustacea</i>	240.00	504	1	2	120.00

Roberts et al. [12]	CNT	NOEC	<i>Daphnia magna, crustacea</i>	5,000.00	96	10	1	500.00
Schwab et al. [7]	CNT	NOEC	<i>Pseudokirchneriella subcapitata</i>	1,300.00	96	1	1	1,300.00
Schwab et al. [7]	CNT	NOEC	<i>Pseudokirchneriella subcapitata</i>	3,000.00	96	1	1	3,000.00
Bayat, et.al. [13]	CNT	LED	<i>Saccharomyces cerevisiae</i>	7,800.00	16	10	2	390.00
Zhu et al. [14]	CNT	NOEC	<i>Stylonychia Mytilus</i>	500.00	120	1	1	500.00
Ghafari et al. [15]	CNT	LOEC	<i>Tetrahymena thermophila, protozoa</i>	3,600.00	72	1	2	1,800.00
Mouchet, et.al. [16]	CNT	LOEC	<i>Xenopus laevis, amphibian</i>	10,000.00	288	10	2	500.00
Mouchet, et.al. [17]	CNT	LOEC	<i>Xenopus laevis, amphibian</i>	50,000.00	288	10	2	2,500.00
Fang, et.al. [18]	Fullerenes	MIC	<i>Bacillus subtilis, bacteria</i>	612.37	14	10	2	30.62
Zhu et al. [19]	Fullerenes	NOEC	<i>Carassius auratus</i>	40.00	768	1	1	40.00
Luo et al. [20]	Fullerenes	LOEC	<i>Chlamydomonas reinhardtii, unicellular</i>	1,000.00	24	10	2	50.00
Isaacson et al. [21]	Fullerenes	LC50	<i>Danio rerio, Zebrafish</i>	130.00	12	10	10	1.30
Usenko et al. [22]	Fullerenes	NOEC	<i>Danio rerio, Zebrafish</i>	100.00	24	10	1	10.00
Seda [23]	Fullerenes	LC50	<i>Daphnia magna, crustacea</i>	400.00	96	10	10	4.00
Tao, et.al. [24]	Fullerenes	LOEC	<i>Daphnia magna, crustacea</i>	200.00	48	10	2	10.00
Lovern et al. [25]	Fullerenes	NOEC	<i>Daphnia magna, crustacea</i>	200.00	48	10	1	20.00
Zhu et al. [26]	Fullerenes	NOEC	<i>Daphnia magna, crustacea</i>	500.00	192	10	1	50.00
Zhu et al. [27]	Fullerenes	EC50	<i>Daphnia magna, crustacea</i>	9,344.00	48	10	10	93.44
Oberdörster et al. [28]	Fullerenes	HONEC	<i>Daphnia magna, crustacea</i>	5,000.00	504	1	1	5,000.00
Ivask, et.al. [29]	Fullerenes	EC50	<i>Escherichia coli</i>	20,800,000.00	2	10	10	208,000.00

Oberdörster, et al. [30]	Fullerenes	LOEC	<i>Micropterus salmoides, Largemouth bass</i>	500.00	48	10	2	25.00
Oberdörster et al. [28]	Fullerenes	HONEC	<i>Oryzias latipes, japanese rice fish</i>	500.00	96	10	1	50.00
Oberdörster et al. [28]	Fullerenes	HONEC	<i>Pimephales promelas, fathead minnow</i>	500.00	96	10	1	50.00
Fang, et al. [18]	Fullerenes	MIC	<i>Pseudomonas putida, bacteria</i>	353.55	14	10	2	17.68
Gao, et al. [31]	Nano-Ag	HONEC	<i>ammonia-oxidizing bacteria (AOB)</i>	200.00	408	1	1	200.00
Luo, et al. [32]	Nano-Ag	HONEC	<i>ammonia-oxidizing bacteria (AOB)</i>	2,000.00	1080	1	1	2,000.00
Jin [33]	Nano-Ag	IC50	<i>Bacillus subtilis, bacteria</i>	3,000.00	24	10	10	30.00
Yoon et al. [34]	Nano-Ag	LC50	<i>Bacillus subtilis, bacteria</i>	11,000.00	24	10	10	110.00
Jin [33]	Nano-Ag	IC50	<i>Bacillus subtilis, bacteria</i>	46,000.00	24	10	10	460.00
Roh et al. [35]	Nano-Ag	LOEC	<i>Caenorhabditis elegans, roundworm</i>	100.00	72	10	2	5.00
Kim et al. [36]	Nano-Ag	NOEC	<i>Caenorhabditis elegans, roundworm</i>	1,000.00	24	10	10	10.00
Meyer et al. [37]	Nano-Ag	LOEC	<i>Caenorhabditis elegans, roundworm</i>	5,000.00	48	10	2	250.00
Kim et al. [36]	Nano-Ag	LC50	<i>Caenorhabditis elegans, roundworm</i>	55,000.00	24	10	10	550.00
Angel [38]	Nano-Ag	IC50	<i>Ceriodaphnia dubia, crustacea</i>	0.15	72	10	10	0.002
Gao et al. [39]	Nano-Ag	LC50	<i>Ceriodaphnia dubia, crustacea</i>	0.69	48	10	10	0.01
Gao et al. [39]	Nano-Ag	LC50	<i>Ceriodaphnia dubia, crustacea</i>	0.77	48	10	10	0.01
Angel [38]	Nano-Ag	IC50	<i>Ceriodaphnia dubia, crustacea</i>	2.00	72	10	10	0.02
Kennedy et al. [40]	Nano-Ag	LC50	<i>Ceriodaphnia dubia, crustacea</i>	3.00	48	10	10	0.03

Kennedy et al. [40]	Nano-Ag	LC50	<i>Ceriodaphnia dubia, crustacea</i>	4.80	48	10	10	0.05
Gao et al. [39]	Nano-Ag	LC50	<i>Ceriodaphnia dubia, crustacea</i>	6.18	48	10	10	0.06
Kennedy et al. [40]	Nano-Ag	LC50	<i>Ceriodaphnia dubia, crustacea</i>	8.80	48	10	10	0.09
Kennedy et al. [40]	Nano-Ag	LC50	<i>Ceriodaphnia dubia, crustacea</i>	10.00	48	10	10	0.10
Kennedy et al. [40]	Nano-Ag	LC50	<i>Ceriodaphnia dubia, crustacea</i>	11.00	48	10	10	0.11
Kennedy et al. [40]	Nano-Ag	LC50	<i>Ceriodaphnia dubia, crustacea</i>	19.00	48	10	10	0.19
Kennedy et al. [40]	Nano-Ag	LC50	<i>Ceriodaphnia dubia, crustacea</i>	26.00	48	10	10	0.26
Kennedy et al. [40]	Nano-Ag	LC50	<i>Ceriodaphnia dubia, crustacea</i>	26.70	48	10	10	0.27
Kennedy et al. [40]	Nano-Ag	LC50	<i>Ceriodaphnia dubia, crustacea</i>	29.00	48	10	10	0.29
Kennedy et al. [40]	Nano-Ag	LC50	<i>Ceriodaphnia dubia, crustacea</i>	38.10	48	10	10	0.38
Kennedy et al. [40]	Nano-Ag	LC50	<i>Ceriodaphnia dubia, crustacea</i>	41.40	48	10	10	0.41
Kennedy et al. [40]	Nano-Ag	LC50	<i>Ceriodaphnia dubia, crustacea</i>	45.00	48	10	10	0.45
Griffit et al. [41]	Nano-Ag	LC50	<i>Ceriodaphnia dubia, crustacea</i>	67.00	48	10	10	0.67
McLaughlin [42]	Nano-Ag	LC50	<i>Ceriodaphnia dubia, crustacea</i>	433.00	48	10	10	4.33
Nair, et.al. [43]	Nano-Ag	HONEC	<i>Chironomus riparius</i>	1,000.00	24	10	1	100.00
Navarro et al. [44]	Nano-Ag	EC50	<i>Chlamydomonas reinhardtii, unicellular</i>	0.86	5	10	10	0.01
Navarro [44]	Nano-Ag	EC50	<i>Chlamydomonas reinhardtii, unicellular</i>	3.53	1	10	10	0.04
Navarro [44]	Nano-Ag	EC50	<i>Chlamydomonas reinhardtii, unicellular</i>	6.10	1	10	10	0.06

Wang et al. [45]	Nano-Ag	EC50	<i>Chydorus sphaericus (cladoceran species)</i>	8.63	48	10	10	0.09
Wang et al. [45]	Nano-Ag	EC50	<i>Chydorus sphaericus (cladoceran species)</i>	12.94	48	10	10	0.13
Wang et al. [45]	Nano-Ag	EC50	<i>Chydorus sphaericus (cladoceran species)</i>	30.20	48	10	10	0.30
Massarsky [46]	Nano-Ag	LC50	<i>Danio rerio, Zebrafish</i>	1.18	96	10	10	0.01
Kaewamatawong [47]	Nano-Ag	LC50	<i>Danio rerio, Zebrafish</i>	1.78	48	10	10	0.02
Cunningham [48]	Nano-Ag	EC50	<i>Danio rerio, Zebrafish</i>	8.45	48	10	10	0.08
Cunningham [48]	Nano-Ag	EC50	<i>Danio rerio, Zebrafish</i>	34.50	48	10	10	0.35
Cunningham [48]	Nano-Ag	EC50	<i>Danio rerio, Zebrafish</i>	45.50	48	10	10	0.46
Cunningham [48]	Nano-Ag	EC50	<i>Danio rerio, Zebrafish</i>	61.00	48	10	10	0.61
Wang et al. [45]	Nano-Ag	EC50	<i>Danio rerio, Zebrafish</i>	84.14	48	10	10	0.84
Bilberg [49]	Nano-Ag	LC50	<i>Danio rerio, Zebrafish</i>	89.00	48	10	10	0.89
Wang et al. [45]	Nano-Ag	EC50	<i>Danio rerio, Zebrafish</i>	146.70	48	10	10	1.47
Wang et al. [45]	Nano-Ag	EC50	<i>Danio rerio, Zebrafish</i>	173.67	48	10	10	1.74
Cunningham [48]	Nano-Ag	EC50	<i>Danio rerio, Zebrafish</i>	205.00	48	10	10	2.05
Cunningham [48]	Nano-Ag	EC50	<i>Danio rerio, Zebrafish</i>	228.00	48	10	10	2.28
Cunningham [48]	Nano-Ag	EC50	<i>Danio rerio, Zebrafish</i>	351.00	48	10	10	3.51
Cunningham [48]	Nano-Ag	EC50	<i>Danio rerio, Zebrafish</i>	370.00	48	10	10	3.70
Muth-Köhne [50]	Nano-Ag	EC50	<i>Danio rerio, Zebrafish</i>	1,000.00	48	10	10	10.00
Muth-Köhne [50]	Nano-Ag	EC50	<i>Danio rerio, Zebrafish</i>	1,200.00	24	10	10	12.00
Muth-Köhne [50]	Nano-Ag	LC50	<i>Danio rerio, Zebrafish</i>	1,200.00	48	10	10	12.00

Muth-Köhne [50]	Nano-Ag	LC50	<i>Danio rerio, Zebrafish</i>	1,900.00	24	10	10	19.00
van Aerle [51]	Nano-Ag	LC50	<i>Danio rerio, Zebrafish</i>	1,912.80	48	10	10	19.13
Cunningham [48]	Nano-Ag	EC50	<i>Danio rerio, Zebrafish</i>	2,427.00	48	10	10	24.27
Cunningham [48]	Nano-Ag	EC50	<i>Danio rerio, Zebrafish</i>	3,043.00	48	10	10	30.43
Cunningham [48]	Nano-Ag	EC50	<i>Danio rerio, Zebrafish</i>	3,091.00	48	10	10	30.91
Cunningham [48]	Nano-Ag	EC50	<i>Danio rerio, Zebrafish</i>	3,455.00	48	10	10	34.55
Cunningham [48]	Nano-Ag	EC50	<i>Danio rerio, Zebrafish</i>	5,891.00	48	10	10	58.91
Cunningham [48]	Nano-Ag	EC50	<i>Danio rerio, Zebrafish</i>	6,922.00	48	10	10	69.22
Griffit et al. [41]	Nano-Ag	LC50	<i>Danio rerio, Zebrafish</i>	7,070.00	48	10	10	70.70
Asharani et al. [52]	Nano-Ag	LC50	<i>Danio rerio, Zebrafish</i>	35,355.34	72	10	10	353.55
Choi, et.al. [53]	Nano-Ag	LC50	<i>Danio rerio, Zebrafish</i>	250,000.00	24	10	10	2,500.00
Völker [54]	Nano-Ag	EC10	<i>Daphnia galeata crustacea</i>	11.00	48	10	2	0.55
Völker [54]	Nano-Ag	EC10	<i>Daphnia galeata crustacea</i>	3.45	504	1	2	1.73
Lee[55]	Nano-Ag	EC50	<i>Daphnia magna, crustacea</i>	0.75	48	10	10	0.01
Kim et al. [56]	Nano-Ag	EC50	<i>Daphnia magna, crustacea</i>	1.00	48	10	10	0.01
Zhao et al. [57]	Nano-Ag	LC50	<i>Daphnia magna, crustacea</i>	1.10	48	10	10	0.01
Allen, et.al. [58]	Nano-Ag	LC50	<i>Daphnia magna, crustacea</i>	1.10	48	10	10	0.01
Georgantzopoulou [59]	Nano-Ag	EC50	<i>Daphnia magna, crustacea</i>	1.20	48	10	10	0.01
Kim et al. [56]	Nano-Ag	EC50	<i>Daphnia magna, crustacea</i>	1.40	48	10	10	0.01

Poynton [60]	Nano-Ag	EC50	<i>Daphnia magna, crustacea</i>	1.80	24	10	10	0.02
Kennedy et al. [61]	Nano-Ag	LC50	<i>Daphnia magna, crustacea</i>	1.80	48	10	10	0.02
Zhao et al. [57]	Nano-Ag	LC50	<i>Daphnia magna, crustacea</i>	2.00	48	10	10	0.02
Asghari [62]	Nano-Ag	EC50	<i>Daphnia magna, crustacea</i>	2.00	48	10	10	0.02
Newton [63]	Nano-Ag	LC50	<i>Daphnia magna, crustacea</i>	2.14	48	10	10	0.02
Newton [63]	Nano-Ag	LC50	<i>Daphnia magna, crustacea</i>	2.27	48	10	10	0.02
Das, et.al. [64]	Nano-Ag	LC50	<i>Daphnia magna, crustacea</i>	2.75	48	10	10	0.03
Newton [63]	Nano-Ag	LC50	<i>Daphnia magna, crustacea</i>	3.16	48	10	10	0.03
Newton [63]	Nano-Ag	LC50	<i>Daphnia magna, crustacea</i>	3.41	48	10	10	0.03
Li et al. [65]	Nano-Ag	LC50	<i>Daphnia magna, crustacea</i>	3.46	48	10	10	0.03
Newton [63]	Nano-Ag	LC50	<i>Daphnia magna, crustacea</i>	3.48	48	10	10	0.03
Li et al. [65]	Nano-Ag	LC50	<i>Daphnia magna, crustacea</i>	4.00	48	10	10	0.04
Asghari [62]	Nano-Ag	EC50	<i>Daphnia magna, crustacea</i>	4.00	48	10	10	0.04
Jo [66]	Nano-Ag	EC50	<i>Daphnia magna, crustacea</i>	4.20	24	10	10	0.04
Kennedy et al. [61]	Nano-Ag	LC50	<i>Daphnia magna, crustacea</i>	5.30	48	10	10	0.05
Kennedy et al. [61]	Nano-Ag	LC50	<i>Daphnia magna, crustacea</i>	5.40	48	10	10	0.05
Kennedy et al. [61]	Nano-Ag	LC50	<i>Daphnia magna, crustacea</i>	5.40	48	10	10	0.05
Li et al. [65]	Nano-Ag	LC50	<i>Daphnia magna, crustacea</i>	6.00	48	10	10	0.06
Hoheisel [67]	Nano-Ag	LC50	<i>Daphnia magna, crustacea</i>	7.00	48	10	10	0.07

Das, et.al. [64]	Nano-Ag	NOEC	<i>Daphnia magna, crustacea</i>	0.75	48	10	1	0.08
Lee [55]	Nano-Ag	EC50	<i>Daphnia magna, crustacea</i>	7.98	48	10	10	0.08
Li et al. [65]	Nano-Ag	LC50	<i>Daphnia magna, crustacea</i>	10.00	48	10	10	0.10
Stensberg [68]	Nano-Ag	LC50	<i>Daphnia magna, crustacea</i>	10.10	72	10	10	0.10
Poynton [60]	Nano-Ag	EC50	<i>Daphnia magna, crustacea</i>	10.60	24	10	10	0.11
Kennedy et al. [61]	Nano-Ag	LC50	<i>Daphnia magna, crustacea</i>	11.10	48	10	10	0.11
Jo [66]	Nano-Ag	EC50	<i>Daphnia magna, crustacea</i>	12.40	24	10	10	0.12
Newton [63]	Nano-Ag	LC50	<i>Daphnia magna, crustacea</i>	13.08	48	10	10	0.13
Newton [63]	Nano-Ag	LC50	<i>Daphnia magna, crustacea</i>	14.04	48	10	10	0.14
Newton [63]	Nano-Ag	LC50	<i>Daphnia magna, crustacea</i>	14.09	48	10	10	0.14
Jo [66]	Nano-Ag	EC50	<i>Daphnia magna, crustacea</i>	14.30	24	10	10	0.14
Newton [63]	Nano-Ag	LC50	<i>Daphnia magna, crustacea</i>	14.81	48	10	10	0.15
Kennedy et al. [61]	Nano-Ag	LC50	<i>Daphnia magna, crustacea</i>	14.90	48	10	10	0.15
Kennedy et al. [61]	Nano-Ag	LC50	<i>Daphnia magna, crustacea</i>	17.70	48	10	10	0.18
Georgantzopoulou [59]	Nano-Ag	EC50	<i>Daphnia magna, crustacea</i>	20.00	48	10	10	0.20
Hoheisel [67]	Nano-Ag	LC50	<i>Daphnia magna, crustacea</i>	27.00	48	10	10	0.27
Zhao et al. [57]	Nano-Ag	LC50	<i>Daphnia magna, crustacea</i>	28.70	48	10	10	0.29
Völker [54]	Nano-Ag	EC10	<i>Daphnia magna, crustacea</i>	0.92	504	1	2	0.46
Kennedy et al. [61]	Nano-Ag	LC50	<i>Daphnia magna, crustacea</i>	97.00	48	10	10	0.97



Georgantzopoulou [59]	Nano-Ag	EC50	<i>Daphnia magna, crustacea</i>	120.00	48	10	10	1.20
Asghari [62]	Nano-Ag	EC50	<i>Daphnia magna, crustacea</i>	187.00	48	10	10	1.87
Zhao et al. [69]	Nano-Ag	LOEC	<i>Daphnia magna, crustacea</i>	5.00	504	1	2	2.50
Völker [54]	Nano-Ag	EC10	<i>Daphnia magna, crustacea</i>	60.30	48	10	2	3.02
Jo [66]	Nano-Ag	EC50	<i>Daphnia magna, crustacea</i>	531.50	24	10	10	5.32
Pokhrel [70]	Nano-Ag	EC50	<i>Daphnia magna, crustacea</i>	798.00	48	10	10	7.98
Jo [66]	Nano-Ag	EC50	<i>Daphnia magna, crustacea</i>	1,153.90	24	10	10	11.54
Jo [66]	Nano-Ag	EC50	<i>Daphnia magna, crustacea</i>	1,404.60	24	10	10	14.05
Jo [66]	Nano-Ag	EC50	<i>Daphnia magna, crustacea</i>	3,844.10	24	10	10	38.44
Zhao et al. [69]	Nano-Ag	HONEC	<i>Daphnia magna, crustacea</i>	500.00	48	10	1	50.00
Blinova et al. [71]	Nano-Ag	EC50	<i>Daphnia magna, crustacea</i>	40,200.00	48	10	10	402.00
Blinova et al. [71]	Nano-Ag	EC50	<i>Daphnia magna, crustacea</i>	49,400.00	48	10	10	494.00
Blinova et al. [71]	Nano-Ag	EC50	<i>Daphnia magna, crustacea</i>	263,300.00	48	10	10	2,633.00
Völker [54]	Nano-Ag	EC10	<i>Daphnia pulex, crustacea</i>	4.37	48	10	2	0.22
Griffit et al.	Nano-Ag	LC50	<i>Daphnia pulex, crustacea</i>	40.00	48	10	10	0.40
Völker [54]	Nano-Ag	EC10	<i>Daphnia pulex, crustacea</i>	2.25	504	1	2	1.13
Georgantzopoulou [59]	Nano-Ag	EC50	<i>Desmodesmus subspicatus</i>	34.00	72	1	10	3.40
Georgantzopoulou [59]	Nano-Ag	EC50	<i>Desmodesmus subspicatus</i>	330.00	72	1	10	33.00
Georgantzopoulou [59]	Nano-Ag	EC50	<i>Desmodesmus subspicatus</i>	2,200.00	72	1	10	220.00
Martinez-Castañon, et.al. [72]	Nano-Ag	MIC	<i>Escherichia coli</i>	6.25	24	10	2	0.31

Pokhrel [73]	Nano-Ag	EC50	<i>Escherichia coli</i>	800.00	5	10	10	8.00
Pokhrel [73]	Nano-Ag	EC50	<i>Escherichia coli</i>	4,170.00	5	10	10	41.70
Pokhrel [73]	Nano-Ag	EC50	<i>Escherichia coli</i>	5,790.00	5	10	10	57.90
Yoon et al. [34]	Nano-Ag	LC50	<i>Escherichia coli</i>	31,000.00	24	10	10	310.00
Ivask, et.al. [29]	Nano-Ag	EC50	<i>Escherichia coli</i>	45,900.00	2	10	10	459.00
Funck, et.al. [74]	Nano-Ag	LC50	<i>Gammarus fossaru, amphipod</i>	1.01	96	10	10	0.01
Gubbins, et.al. [75]	Nano-Ag	EC50	<i>Lemna minor, duck weed</i>	19.06	336	10	10	0.19
Kim et al. [76]	Nano-Ag	NOEC	<i>Lemna paucicostata, duck weed</i>	100.00	168	10	1	10.00
Ali, et.al. [77]	Nano-Ag	LC50	<i>Lymnaea luteola, snail</i>	48.10	96	10	10	0.48
Miao [78]	Nano-Ag	EC50	<i>Ochromonas danica, chrysophyte</i>	5.30	48	10	10	0.05
Govindasamy, et al [79]	Nano-Ag	LC50	<i>Oreochromis mossambicus, tilapia</i>	12,600.00	192	10	10	126.00
Kim et al. [56]	Nano-Ag	LC50	<i>Oryzias latipes, japanese rice fish</i>	28.00	96	10	10	0.28
Chae [80]	Nano-Ag	LC50	<i>Oryzias latipes, japanese rice fish</i>	34.60	96	10	10	0.35
Kim et al. [56]	Nano-Ag	LC50	<i>Oryzias latipes, japanese rice fish</i>	67.00	96	10	10	0.67
Wu, et.al. [81]	Nano-Ag	LC50	<i>Oryzias latipes, japanese rice fish</i>	1,030.00	48	10	10	10.30
Kashiwada, et.al. [82]	Nano-Ag	LC50	<i>Oryzias latipes, japanese rice fish</i>	1,390.00	96	10	10	13.90
Kim et al. [83]	Nano-Ag	LC50	<i>Oryzias latipes, japanese rice fish</i>	1,440.00	96	10	10	14.40
Kvitek et al. [84]	Nano-Ag	LC50	<i>Paramecium caudatum</i>	39,000.00	1	10	10	390.00
Kvitek et al. [84]	Nano-Ag	NOEC	<i>Paramecium caudatum</i>	25,000.00	168	1	1	25,000.00
Bilberg [85]	Nano-Ag	HONEC	<i>Perca fluviatilis</i>	300.00	20	10	1	30.00
Angel [38]	Nano-Ag	IC50	<i>Phaeodactylum tricornutum</i>	2,380.00	72	1	10	238.00

Angel [38]	Nano-Ag	IC50	<i>Phaeodactylum tricornutum</i>	3,690.00	72	1	10	369.00
Kennedy et al. [61]	Nano-Ag	LC50	<i>Pimephales promelas, fathead minnow</i>	9.00	48	10	10	0.09
Kennedy et al. [61]	Nano-Ag	LC50	<i>Pimephales promelas, fathead minnow</i>	19.20	48	10	10	0.19
Kennedy et al. [61]	Nano-Ag	LC50	<i>Pimephales promelas, fathead minnow</i>	41.00	48	10	10	0.41
Kennedy et al. [61]	Nano-Ag	LC50	<i>Pimephales promelas, fathead minnow</i>	55.20	48	10	10	0.55
Kennedy et al. [61]	Nano-Ag	LC50	<i>Pimephales promelas, fathead minnow</i>	60.70	48	10	10	0.61
Kennedy et al. [61]	Nano-Ag	LC50	<i>Pimephales promelas, fathead minnow</i>	64.10	48	10	10	0.64
Kennedy et al. [61]	Nano-Ag	LC50	<i>Pimephales promelas, fathead minnow</i>	69.90	48	10	10	0.70
Hoheisel [67]	Nano-Ag	LC50	<i>Pimephales promelas, fathead minnow</i>	89.40	96	10	10	0.89
Kennedy et al. [61]	Nano-Ag	LC50	<i>Pimephales promelas, fathead minnow</i>	125.80	48	10	10	1.26
Hoheisel [67]	Nano-Ag	EC20	<i>Pimephales promelas, fathead minnow</i>	46.10	168	10	2	2.31
Laban et al. [86]	Nano-Ag	LC50	<i>Pimephales promelas, fathead minnow</i>	1,250.00	96	10	10	12.50
Laban et al. [86]	Nano-Ag	LC50	<i>Pimephales promelas, fathead minnow</i>	1,303.84	96	10	10	13.04
Laban et al. [86]	Nano-Ag	LC50	<i>Pimephales promelas, fathead minnow</i>	1,360.00	96	10	10	13.60
Laban et al. [86]	Nano-Ag	LC50	<i>Pimephales promelas, fathead minnow</i>	9,400.00	96	10	10	94.00
Laban et al. [86]	Nano-Ag	LC50	<i>Pimephales promelas, fathead minnow</i>	9,981.98	96	10	10	99.82
Laban et al. [86]	Nano-Ag	LC50	<i>Pimephales promelas, fathead minnow</i>	10,600.00	96	10	10	106.00
Angel [38]	Nano-Ag	IC50	<i>Pseudokirchneriella subcapitata</i>	3.00	72	1	10	0.30
Angel [38]	Nano-Ag	IC50	<i>Pseudokirchneriella subcapitata</i>	19.50	72	1	10	1.95
Griffit et al. [41]	Nano-Ag	EC50	<i>Pseudokirchneriella subcapitata</i>	190.00	96	1	10	19.00
McLaughlin [42]	Nano-Ag	IC50	<i>Pseudokirchneriella subcapitata</i>	22,600.00	96	1	1	22,600.00
Jin [33]	Nano-Ag	IC50	<i>Pseudomonas putida, bacteria</i>	3,000.00	24	10	10	30.00
Jin [33]	Nano-Ag	IC50	<i>Pseudomonas putida, bacteria</i>	50,000.00	24	10	10	500.00
Wang et al. [45]	Nano-Ag	EC50	<i>Raphidocelis subcapitata</i>	896.39	4.5	10	10	8.96
Wang et al. [45]	Nano-Ag	EC50	<i>Raphidocelis subcapitata</i>	4,006.23	4.5	10	10	40.06
Wang et al. [45]	Nano-Ag	EC50	<i>Raphidocelis subcapitata</i>	21,142.19	4.5	10	10	211.42
Debabrata, et.al. [87]	Nano-Ag	MIC	<i>Saccharomyces cerevisiae</i>	48.51		10	2	2.43
Niazi, et.al. [88]	Nano-Ag	EC30	<i>Saccharomyces cerevisiae</i>	15,101.56	2	10	2	755.08

Bayat, et.al. [13]	Nano-Ag	LED	<i>Saccharomyces cerevisiae</i>	15,600.00	16	10	2	780.00
Jiang [89]	Nano-Ag	EC50	<i>Spirodela polyrhiza, duckweed</i>	4,540.00	72	10	10	45.40
Jiang [89]	Nano-Ag	EC50	<i>Spirodela polyrhiza, duckweed</i>	13,390.00	72	10	10	133.90
Jiang [89]	Nano-Ag	EC50	<i>Spirodela polyrhiza, duckweed</i>	13,670.00	72	10	10	136.70
Jiang [89]	Nano-Ag	EC50	<i>Spirodela polyrhiza, duckweed</i>	16,100.00	72	10	10	161.00
Jiang [89]	Nano-Ag	EC50	<i>Spirodela polyrhiza, duckweed</i>	17,330.00	72	10	10	173.30
Martinez-Castañon, et.al. [72]	Nano-Ag	MIC	<i>Staphylococcus aureus</i>	7.50	24	10	2	0.38
Burchardt[90]	Nano-Ag	EC50	<i>Synechococcus, bacteria sp.</i>	355.97	72	1	10	35.60
Juganson [91]	Nano-Ag	EC50	<i>Tetrahymena thermophila, protozoa</i>	205,000.00	24	10	10	2,050.00
Juganson [91]	Nano-Ag	EC50	<i>Tetrahymena thermophila, protozoa</i>	286,000.00	2	10	10	2,860.00
Blinova et al. [71]	Nano-Ag	LC50	<i>Thamnocephalus platyurus</i>	68,800.00	48	10	10	688.00
Blinova et al. [71]	Nano-Ag	LC50	<i>Thamnocephalus platyurus</i>	178,000.00	48	10	10	1,780.00
Blinova et al. [71]	Nano-Ag	LC50	<i>Thamnocephalus platyurus</i>	191,500.00	48	10	10	1,915.00
Blinova et al. [71]	Nano-Ag	LC50	<i>Thamnocephalus platyurus</i>	250,000.00	48	10	10	2,500.00
Blinova et al. [71]	Nano-Ag	LC50	<i>Thamnocephalus platyurus</i>	252,000.00	48	10	10	2,520.00
Cherchi et al. [92]	Nano-TiO2	EC50	<i>Anabaena variabilis</i>	620.00	96	1	10	62.00
Adams et al. [93]	Nano-TiO2	NOEC	<i>Bacillus subtilis, bacteria</i>	500,000.00	14 - 20	10	1	50,000.00
Clement, et.al. [94]	Nano-TiO2	EC50	<i>Brachionus plicatili, rotifer</i>	5,370.00	72	10	10	53.70
Wu, et.al. [95]	Nano-TiO2	LOEC	<i>Caenorhabditis elegans, roundworm</i>	50.00	24	10	2	2.50
Wang et al. [96]	Nano-TiO2	LC50	<i>Caenorhabditis elegans, roundworm</i>	80,000.00	24	10	10	800.00
Hall et al. [97]	Nano-TiO2	LC50	<i>Ceriodaphnia dubia, crustacea</i>	7,600.00	48	10	10	76.00
Hall et al. [97]	Nano-TiO2	IC25	<i>Ceriodaphnia dubia, crustacea</i>	8,500.00	168	10	10	85.00
Li et al. [3]	Nano-TiO2	EC50	<i>Ceriodaphnia dubia, crustacea</i>	42,000.00	48	10	10	420.00
Wang et al. [98]	Nano-TiO2	LC50	<i>Ceriodaphnia dubia, crustacea</i>	400,000.00	24	10	10	4,000.00
Li et al. [99]	Nano-TiO2	LC50	<i>Chironomus dilutus</i>	20,000.00	48	10	10	200.00
Wang et al. [100]	Nano-TiO2	NOEC	<i>Chlamydomonas reinhardtii, unicellular</i>	1,000.00	120	1	1	1,000.00
Gunawan, et.al. [101]	Nano-TiO2	EC50	<i>Chlamydomonas reinhardtii, unicellular</i>	100,000.00	192	1	10	10,000.00
Sadiq et al. [102]	Nano-TiO2	EC50	<i>Chlorella sp.</i>	16,120.00	72	1	10	1,612.00

Ji et al. [103]	Nano-TiO2	NOEC	<i>Chlorella sp.</i>	16,000.00	144	1	1	16,000.00
Cheng et al. [104]	Nano-TiO2	HONEC	<i>Danio rerio, Zebrafish</i>	500.00	4320	1	1	500.00
Xiong et al. [105]	Nano-TiO2	LC50	<i>Danio rerio, Zebrafish</i>	124,500.00	96	10	10	1,245.00
Yang [106]	Nano-TiO2	LC50	<i>Danio rerio, Zebrafish</i>	156,000.00	24	10	10	1,560.00
Bar-Ilan [107]	Nano-TiO2	LC50	<i>Danio rerio, Zebrafish</i>	300,000.00	24	10	10	3,000.00
Zhu et al. [108]	Nano-TiO2	HONEC	<i>Danio rerio, Zebrafish</i>	500,000.00	96	10	1	50,000.00
Ma, et.al [109]	Nano-TiO2	LC50	<i>Daphnia magna, crustacea</i>	29.80	48	10	10	0.30
Dabrunz et al. [110]	Nano-TiO2	EC50	<i>Daphnia magna, crustacea</i>	240.00	96	10	10	2.40
Zhu et al. [111]	Nano-TiO2	NOEC	<i>Daphnia magna, crustacea</i>	100.00	72	10	1	10.00
Clement, et.al. [94]	Nano-TiO2	EC50	<i>Daphnia magna, crustacea</i>	1,300.00	72	10	10	13.00
Dabrunz et al. [110]	Nano-TiO2	EC50	<i>Daphnia magna, crustacea</i>	3,800.00	72	10	10	38.00
Das, et.al. [64]	Nano-TiO2	LC50	<i>Daphnia magna, crustacea</i>	7,750.00	48	10	10	77.50
Zhu et al. [111]	Nano-TiO2	NOEC	<i>Daphnia magna, crustacea</i>	100.00	504	1	1	100.00
Warheit et al. [112]	Nano-TiO2	NOEC	<i>Daphnia magna, crustacea</i>	1,000.00	48	10	1	100.00
Lovern 2006[25]	Nano-TiO2	NOEC	<i>Daphnia magna, crustacea</i>	1,000.00	48	10	1	100.00
Kim et al. [113]	Nano-TiO2	LOEC	<i>Daphnia magna, crustacea</i>	500.00	504	1	2	250.00
Kim et al. [113]	Nano-TiO2	LOEC	<i>Daphnia magna, crustacea</i>	5,000.00	48	10	2	250.00
Jacobasch, et.al. [114]	Nano-TiO2	EC50	<i>Daphnia magna, crustacea</i>	2,930.00	504	1	10	293.00
Amiano [115]	Nano-TiO2	EC50	<i>Daphnia magna, crustacea</i>	29,700.00	48	10	10	297.00
Zhu et al. [27]	Nano-TiO2	EC50	<i>Daphnia magna, crustacea</i>	35,306.00	48	10	10	353.06
Das, et.al. [64]	Nano-TiO2	NOEC	<i>Daphnia magna, crustacea</i>	4,100.00	48	10	1	410.00
Jacobasch, et.al. [114]	Nano-TiO2	EC10	<i>Daphnia magna, crustacea</i>	4,520.00	504	1	2	2,260.00
Wiench et al. [116]	Nano-TiO2	NOEC	<i>Daphnia magna, crustacea</i>	3,000.00	504	1	1	3,000.00
Heinlaan et al. [117]	Nano-TiO2	LC50	<i>Daphnia magna, crustacea</i>	20,000,000.00	48	10	10	200,000.00
Hall et al. [97]	Nano-TiO2	LC50	<i>Daphnia pulex, crustacea</i>	9,200.00	48	10	10	92.00
Marcone [118]	Nano-TiO2	EC50	<i>Daphnia similis, crustacea</i>	12,500.00	48	10	10	125.00
Hund-Rinke et al. [119]	Nano-TiO2	EC50	<i>Desmodesmus subspicatus</i>	44,000.00	72	1	10	4,400.00
Bigorgne, et.al. [120]	Nano-TiO2	HONEC	<i>Eisenia fetida, earthworm</i>	10,000.00	24	10	1	1,000.00

Dasari, et.al. [121]	Nano-TiO2	LC50	<i>Escherichia coli</i>	1,680.00	0.5	10	10	16.80
Ivask, et.al. [29]	Nano-TiO2	EC50	<i>Escherichia coli</i>	20,000,000.00	2	10	10	200,000.00
Adams et al. [93]	Nano-TiO2	NOEC	<i>Escherichia coli</i>	100,000.00	14 - 20	10	1	10,000.00
Hu et al. [122]	Nano-TiO2	LD50	<i>Escherichia coli</i>	1,104,000.00	2	10	10	11,040.00
Bundschuh, et.al. [123]	Nano-TiO2	LOEC	<i>Gammarus fossaru, amphipod</i>	200.00	168	10	2	10.00
Zhu et al. [124]	Nano-TiO2	NOEC	<i>Haliotis diversicolor supertexta</i>	2,000.00	10	10	1	200.00
Zhu et al. [124]	Nano-TiO2	EC50	<i>Haliotis diversicolor supertexta</i>	56,900.00	10	10	10	569.00
Li et al. [99]	Nano-TiO2	LC50	<i>Hyalella azteca, amphipod</i>	20,000.00	48	10	10	200.00
Li et al. [125]	Nano-TiO2	LC50	<i>Hyalella azteca, amphipod</i>	631,000.00	96	10	10	6,310.00
Kim et al. [76]	Nano-TiO2	NOEC	<i>Lemna paucicostata, duck weed</i>	125,000.00	168	10	1	12,500.00
Li et al. [99]	Nano-TiO2	LC50	<i>Lumbriculus variegatus</i>	20,000.00	48	10	10	200.00
Federici, et. Al. [126]	Nano-TiO2	HONEC	<i>Oncorhynchus mykiss</i>	1,000.00	168	10	1	100.00
Warheit et al. [112]	Nano-TiO2	NOEC	<i>Oncorhynchus mykiss</i>	1,000.00	96	10	1	100.00
Ma, et.al[109]	Nano-TiO2	LC50	<i>Oryzias latipes, japanese rice fish</i>	2,460.00	96	10	10	24.60
Li et al. [127]	Nano-TiO2	LC50	<i>Paramecium multi-micronucleatum</i>	7,215,200.00	48	1	10	721,520.00
Clement, et.al. [94]	Nano-TiO2	EC50	<i>Phaeodactylum tricornutum</i>	10,910.00	72	1	10	1,091.00
Hall et al. [97]	Nano-TiO2	IC25	<i>Pimephales promelas, fathead minnow</i>	452,000.00	168	10	10	4,520.00
Hall et al. [97]	Nano-TiO2	LC50	<i>Pimephales promelas, fathead minnow</i>	500,000.00	48	10	10	5,000.00
Jovanovic, et.al. [128]	Nano-TiO2	HONEC	<i>Pimephales promelas, fathead minnow</i>	1,000,000.00	168	10	1	100,000.00
Lee [129]	Nano-TiO2	NOEC	<i>Pseudokirchneriella subcapitata</i>	50.00	72	1	1	50.00
Lee [129]	Nano-TiO2	EC50	<i>Pseudokirchneriella subcapitata</i>	2,530.00	72	1	10	253.00
Aruoja et al. [130]	Nano-TiO2	NOEC	<i>Pseudokirchneriella subcapitata</i>	984.00	72	1	1	984.00
Hall et al. [97]	Nano-TiO2	IC25	<i>Pseudokirchneriella subcapitata</i>	1,500.00	96	1	1	1,500.00
Warheit et al. [112]	Nano-TiO2	EC50	<i>Pseudokirchneriella subcapitata</i>	16,000.00	72	1	10	1,600.00
Hartmann et al. [131]	Nano-TiO2	EC50	<i>Pseudokirchneriella subcapitata</i>	71,000.00	72	1	10	7,100.00
Metzler et al. [132]	Nano-TiO2	EC50	<i>Pseudokirchneriella subcapitata</i>	113,000.00	96	1	10	11,300.00
Bayat, et.al. [13]	Nano-TiO2	LED	<i>Saccharomyces cerevisiae</i>	7,800.00	16	10	2	390.00
Kasemets [133]	Nano-TiO2	HONEC	<i>Saccharomyces cerevisiae</i>	20,000,000.00	24	10	1	2,000,000.00

Sadiq et al. [102]	Nano-TiO <sub>2</sub>	EC50	<i>Scenedesmus subspicatus</i>	21,200.00	72	1	10	2,120.00
Velzeboer, et. Al. [134]	Nano-TiO <sub>2</sub>	HONEC	<i>soil bacteria</i>	100,000.00	168	1	1	100,000.00
Heinlaan et al. [117]	Nano-TiO <sub>2</sub>	NOEC	<i>Thamnocephalus platyurus</i>	20,000,000.00	24	10	1	2,000,000.00
Nations, et.al. [135]	Nano-TiO <sub>2</sub>	EC10	<i>Xenopus laevis, amphibian</i>	1,000,000.00	96	10	2	50,000.00
Li et al. [136]	Nano-ZnO	IC50	<i>Bacillus subtilis, bacteria</i>	280.00	24	10	10	2.80
Li et al. [136]	Nano-ZnO	IC50	<i>Bacillus subtilis, bacteria</i>	310.00	24	10	10	3.10
Li et al. [136]	Nano-ZnO	IC50	<i>Bacillus subtilis, bacteria</i>	3,300.00	24	10	10	33.00
Jones et al. [137]	Nano-ZnO	EC50	<i>Bacillus subtilis, bacteria</i>	188,000.00	10	10	10	1,880.00
Wu, et.al. [95]	Nano-ZnO	LOEC	<i>Caenorhabditis elegans, roundworm</i>	50.00	24	10	2	2.50
Wang et al. [96]	Nano-ZnO	LC50	<i>Caenorhabditis elegans, roundworm</i>	2,300.00	24	10	10	23.00
Ma et al. [138]	Nano-ZnO	EC50	<i>Caenorhabditis elegans, roundworm</i>	46,000.00	24	10	10	460.00
Ma et al. [139]	Nano-ZnO	LC50	<i>Caenorhabditis elegans, roundworm</i>	70,000.00	24	10	10	700.00
Gunawan, et.al. [101]	Nano-ZnO	EC50	<i>Chlamydomonas reinhardtii, unicellular</i>	10.00	192	1	10	1.00
Luo et al. [20]	Nano-ZnO	LOEC	<i>Chlamydomonas reinhardtii, unicellular</i>	1,000.00	24	10	2	50.00
Lin [140]	Nano-ZnO	IC50	<i>Chlorella sp.</i>	4,900.00	288	1	10	490.00
Ji et al. [103]	Nano-ZnO	NOEC	<i>Chlorella sp.</i>	5,000.00	144	1	1	5,000.00
Fabrega, et.al. [141]	Nano-ZnO	LOEC	<i>Corophium volutator</i>	200.00	2400	1	2	100.00
Zhu et al. [108]	Nano-ZnO	LC50	<i>Danio rerio, Zebrafish</i>	1,800.00	96	10	10	18.00
Yu [142]	Nano-ZnO	LC50	<i>Danio rerio, Zebrafish</i>	3,969.00	96	10	10	39.69
Xiong et al. [105]	Nano-ZnO	LC50	<i>Danio rerio, Zebrafish</i>	4,920.00	96	10	10	49.20
Zhao et al. [143]	Nano-ZnO	NOEC	<i>Daphnia magna, crustacea</i>	0.80	504	1	1	0.80
Zhu et al. [10]	Nano-ZnO	EC50	<i>Daphnia magna, crustacea</i>	622.00	48	10	10	6.22
Wiench et al. [116]	Nano-ZnO	EC50	<i>Daphnia magna, crustacea</i>	1,000.00	48	10	10	10.00
Lopes, et.al. [144]	Nano-ZnO	EC50	<i>Daphnia magna, crustacea</i>	125.00	504	1	10	12.50
Blinova et al. [145]	Nano-ZnO	EC50	<i>Daphnia magna, crustacea</i>	1,700.00	48	10	10	17.00
Naddafi, et.al. [146]	Nano-ZnO	LC50	<i>Daphnia magna, crustacea</i>	2,100.00	48	10	10	21.00
Blinova et al. [145]	Nano-ZnO	EC50	<i>Daphnia magna, crustacea</i>	2,600.00	48	10	10	26.00
Blinova et al. [145]	Nano-ZnO	EC50	<i>Daphnia magna, crustacea</i>	2,800.00	48	10	10	28.00

Heinlaan et al. [117]	Nano-ZnO	EC50	<i>Daphnia magna, crustacea</i>	3,200.00	48	10	10	32.00
Blinova et al. [145]	Nano-ZnO	EC50	<i>Daphnia magna, crustacea</i>	3,300.00	48	10	10	33.00
Blinova et al. [145]	Nano-ZnO	EC50	<i>Daphnia magna, crustacea</i>	3,400.00	48	10	10	34.00
Blinova et al. [145]	Nano-ZnO	EC50	<i>Daphnia magna, crustacea</i>	3,500.00	48	10	10	35.00
Heinlaan et al. [117]	Nano-ZnO	NOEC	<i>Daphnia magna, crustacea</i>	500.00	48	10	1	50.00
Blinova et al. [145]	Nano-ZnO	EC50	<i>Daphnia magna, crustacea</i>	9,000.00	48	10	10	90.00
Poynton et al. [147]	Nano-ZnO	LC10	<i>Daphnia magna, crustacea</i>	3,700.00	24	10	2	185.00
Dasari, et.al. [121]	Nano-ZnO	LC50	<i>Escherichia coli</i>	48.00	0.5	10	10	0.48
Emami-Karvani, et.al. [148]	Nano-ZnO	MIC	<i>Escherichia coli</i>	1,000.00	24	10	2	50.00
Li et al. [136]	Nano-ZnO	IC50	<i>Escherichia coli</i>	17,000.00	12	10	10	170.00
Hu et al. [122]	Nano-ZnO	LD50	<i>Escherichia coli</i>	21,100.00	2	10	10	211.00
Li et al. [136]	Nano-ZnO	IC50	<i>Escherichia coli</i>	34,000.00	12	10	10	340.00
Pokhrel[73]	Nano-ZnO	EC50	<i>Escherichia coli</i>	57,700.00	5	10	10	577.00
Ivask, et.al. [29]	Nano-ZnO	EC50	<i>Escherichia coli</i>	67,200.00	2	10	10	672.00
Adams et al. [93]	Nano-ZnO	EC50	<i>Escherichia coli</i>	1,240,000.00	14-20	10	10	12,400.00
Poynton et al. [149]	Nano-ZnO	LC50	<i>Hyalella azteca, amphipod</i>	78.50	96	10	10	0.79
Miller et al. [150]	Nano-ZnO	NOEC	<i>Isochrysis galbana</i>	500.00	96	1	1	500.00
Lin [151]	Nano-ZnO	IC50	<i>Lolium perenne (ryegrass)</i>	20,000.00	120	10	10	200.00
Lin [152]	Nano-ZnO	IC50	<i>Lolium perenne (ryegrass)</i>	51,000.00	288	10	10	510.00
Li et al. [127]	Nano-ZnO	LC50	<i>Paramecium multi-micronucleatum</i>	573,800.00	48	1	10	57,380.00
Franklin et al. [153]	Nano-ZnO	IC50	<i>Pseudokirchneriella subcapitata</i>	49.00	72	1	10	4.90
Aruoja et al. [130]	Nano-ZnO	NOEC	<i>Pseudokirchneriella subcapitata</i>	17.00	72	1	1	17.00
Lee [154]	Nano-ZnO	NOEC	<i>Pseudokirchneriella subcapitata</i>	50.00	72	1	1	50.00
Lin [151]	Nano-ZnO	IC50	<i>Raphanus sativus, radish</i>	50,000.00	120	10	10	500.00
Kasemets [133]	Nano-ZnO	EC50	<i>Saccharomyces cerevisiae</i>	143,868.00	24	10	10	1,438.68
Emami-Karvani, et.al. [148]	Nano-ZnO	MIC	<i>Staphylococcus aureus</i>	500.00	24	10	2	25.00
Jones et al. [137]	Nano-ZnO	MIC	<i>Staphylococcus aureus</i>	80,000.00	24	10	2	4,000.00



Jones et al. [137]	Nano-ZnO	MIC	<i>Staphylococcus aureus</i>	1,200,000.00	24	10	2	60,000.00
Mortimer et al. [155]	Nano-ZnO	EC50	<i>Tetrahymena thermophila</i> , <i>protozoa</i>	4,300.00	4	10	10	43.00
Mortimer et al. [155]	Nano-ZnO	EC50	<i>Tetrahymena thermophila</i> , <i>protozoa</i>	6,800.00	24	10	10	68.00
Blinova et al. [145]	Nano-ZnO	EC50	<i>Tetrahymena thermophila</i> , <i>protozoa</i>	9,400.00	24	10	10	94.00
Blinova et al. [145]	Nano-ZnO	EC50	<i>Tetrahymena thermophila</i> , <i>protozoa</i>	12,400.00	24	10	10	124.00
Blinova et al. [71]	Nano-ZnO	EC50	<i>Tetrahymena thermophila</i> , <i>protozoa</i>	26,500.00	24	10	10	265.00
Miller et al. [150]	Nano-ZnO	NOEC	<i>Thalassiosira pseudonana</i>	100.00	96	1	1	100.00
Wong [156]	Nano-ZnO	LC50	<i>Thalassiosira pseudonana</i>	3,280.00	96	1	10	328.00
Jarvis [157]	Nano-ZnO	NOEC	<i>Thalassiosira weissflogii</i>	10.00	72	1	1	10.00
Blinova et al. [145]	Nano-ZnO	LC50	<i>Thamnocephalus platyurus</i>	140.00	24	10	10	1.40
Heinlaan et al. [117]	Nano-ZnO	NOEC	<i>Thamnocephalus platyurus</i>	30.00	24	10	1	3.00
Blinova et al. [145]	Nano-ZnO	LC50	<i>Thamnocephalus platyurus</i>	1,100.00	24	10	10	11.00
Blinova et al. [145]	Nano-ZnO	LC50	<i>Thamnocephalus platyurus</i>	1,400.00	24	10	10	14.00
Blinova et al. [145]	Nano-ZnO	LC50	<i>Thamnocephalus platyurus</i>	1,500.00	24	10	10	15.00
Blinova et al. [145]	Nano-ZnO	LC50	<i>Thamnocephalus platyurus</i>	3,600.00	24	10	10	36.00
Blinova et al. [145]	Nano-ZnO	LC50	<i>Thamnocephalus platyurus</i>	5,300.00	24	10	10	53.00
Blinova et al. [145]	Nano-ZnO	LC50	<i>Thamnocephalus platyurus</i>	6,000.00	24	10	10	60.00
Nations, et.al. [135]	Nano-ZnO	EC10	<i>Xenopus laevis</i> , <i>amphibian</i>	1,300.00	96	10	2	65.00

Table S2: Data for soil toxicity for nano-Ag, nano-TiO<sub>2</sub>, nano-ZnO, CNT and Fullerenes.

Author	ENM	Dose descriptor	Test organism	Concentration (µg/kg)	Exposure time (h)	AF time	AF no-effect	Species sensitivity (µg/kg)
De La Torre-Roche [158]	CNT	HONEC	<i>Curcubita pepo, zucchini</i>	5,000,000.00	672	1	1	5,000,000.00
Scott-Fordsmand et al. [159]	CNT	EC50	<i>Eisenia veneta, earthworm</i>	176,000.00	672	1	10	17,600.00
De La Torre-Roche [158]	CNT	LOEC	<i>Glycine max, soy bean</i>	1,000,000.00	672	1	2	500,000.00
Chung et al. [160]	CNT	LOEC	microbial biomass (organic C and N)	500,000.00	480	1	2	250,000.00
Chung, et al. [160]	CNT	HONEC	soil microbial community	50,000.00	264	1	1	50,000.00
Shrestha, et al. [161]	CNT	HONEC	soil microbial community	1,000,000.00	2160	1	1	1,000,000.00
De La Torre-Roche [158]	CNT	HONEC	<i>Solanum lycopersicu, tomato</i>	5,000,000.00	672	1	1	5,000,000.00
De La Torre-Roche [158]	CNT	LOEC	<i>Zea may, maize</i>	500,000.00	672	1	2	250,000.00
De La Torre-Roche [158]	Fullerenes	HONEC	<i>Curcubita pepo, zucchini</i>	5,000,000.00	672	1	1	5,000,000.00
Scott-Fordsmand et al. [159]	Fullerenes	HONEC	<i>Eisenia veneta, earthworm</i>	1,000,000.00	672	1	1	1,000,000.00
Johansen et al. [162]	Fullerenes	EC10	fast growing soil microbacteria	50,000.00	3	10	2	2,500.00
De La Torre-Roche [158]	Fullerenes	LOEC	<i>Glycine max, soy bean</i>	500,000.00	672	1	2	250,000.00
Van der Ploeg et al. [163]	Fullerenes	NOEC	<i>Lumbriculus rubellus (earthworm)</i>	15,400.00	7824	1	1	15,400.00
Johansen et al. [162]	Fullerenes	EC40	slow growing soil microbacteria	50,000.00	336	1	10	5,000.00
Tong et al. [164]	Fullerenes	HONEC	soil microbial community	1,000,000.00	720	1	1	1,000,000.00
De La Torre-Roche [158]	Fullerenes	HONEC	<i>Solanum lycopersicu, tomato</i>	5,000,000.00	672	1	1	5,000,000.00
De La Torre-Roche [158]	Fullerenes	LOEC	<i>Zea may, maize</i>	500,000.00	672	1	2	250,000.00

Schlich [165]	Nano-Ag	NOEC	<i>Eisenia fetida, earthworm</i>	60,000.00	1344	1	1	60,000.00
Shoultz-Wilson et al. [166]	Nano-Ag	NOEC	<i>Eisenia fetida, earthworm</i>	79,450.00	672	1	1	79,450.00
Shoultz-Wilson et al. [166]	Nano-Ag	NOEC	<i>Eisenia fetida, earthworm</i>	84,150.00	672	1	1	84,150.00
Gomes, et al. [167]	Nano-Ag	NOEC	<i>Enchytraeus albidus, Oligochaete</i>	100,000.00	1008	1	1	100,000.00
Lee [168]	Nano-Ag	NOEC	<i>Phaseolus radiatus</i>	100,000.00	120	10	1	10,000.00
Lee [168]	Nano-Ag	NOEC	<i>Sorghum bicolor</i>	2,000,000.00	120	10	1	200,000.00
Heckmann [169]	Nano-TiO2	EC50	<i>Eisenia fetida, earthworm</i>	1,000,000.00	672	1	10	100,000.00
Hu, et al. [170]	Nano-TiO2	LOEC	<i>Eisenia fetida, earthworm</i>	5,000,000.00	168	10	2	250,000.00
Canas, et al. [171]	Nano-TiO2	HONEC	<i>Eisenia fetida, earthworm</i>	10,000,000.00	336	10	1	1,000,000.00
Nogueira, et al. [172]	Nano-TiO2	LOEC	<i>soil microbial community</i>	5,000,000.00	720	1	2	2,500,000.00
Hu, et al. [170]	Nano-ZnO	LOEC	<i>Eisenia fetida, earthworm</i>	1,000,000.00	168	10	2	50,000.00
Heggelund [173]	Nano-ZnO	EC50	<i>Eisenia fetida, earthworm</i>	901,000.00	672	1	10	90,100.00
Heggelund [173]	Nano-ZnO	EC50	<i>Eisenia fetida, earthworm</i>	918,600.00	672	1	10	91,860.00
Heggelund [173]	Nano-ZnO	EC50	<i>Eisenia fetida, earthworm</i>	2,874,000.00	672	1	10	287,400.00
Canas, et al. [171]	Nano-ZnO	HONEC	<i>Eisenia fetida, earthworm</i>	10,000,000.00	336	10	1	1,000,000.00
Hooper et al. [174]	Nano-ZnO	EC50	<i>Eisenia veneta, earthworm</i>	750,000.00	504	10	10	7,500.00
Waalewijn-Kool [175]	Nano-ZnO	EC50	<i>Folsomia candida, springtail</i>	553,000.00	672	1	10	55,300.00
Waalewijn-Kool [176]	Nano-ZnO	EC50	<i>Folsomia candida, springtail</i>	749,000.00	2160	1	10	74,900.00
Waalewijn-Kool [176]	Nano-ZnO	EC50	<i>Folsomia candida, springtail</i>	873,000.00	672	1	10	87,300.00
Waalewijn-Kool [175]	Nano-ZnO	EC50	<i>Folsomia candida, springtail</i>	1,481,000.00	672	1	10	148,100.00
Waalewijn-Kool [176]	Nano-ZnO	EC50	<i>Folsomia candida, springtail</i>	1,817,000.00	8760	1	10	181,700.00

Waalewijn-Kool [176]	Nano-ZnO	EC50	<i>Folsomia candida, springtail</i>	1,964,000.00	672	1	10	196,400.00
Waalewijn-Kool [176]	Nano-ZnO	EC50	<i>Folsomia candida, springtail</i>	2,847,000.00	2160	1	10	284,700.00
Waalewijn-Kool [175]	Nano-ZnO	EC50	<i>Folsomia candida, springtail</i>	3,233,000.00	672	1	10	323,300.00
Waalewijn-Kool [176]	Nano-ZnO	EC50	<i>Folsomia candida, springtail</i>	5,855,000.00	8760	1	10	585,500.00
Kool [177]	Nano-ZnO	EC10	<i>Folsomia candida, springtail</i>	1,678,000.00	672	1	2	839,000.00
Manzo et al. [178]	Nano-ZnO	EC25	<i>Lepidium sativum (garden cress)</i>	286,000.00	72	10	10	2,860.00
Tourinho, et.al. [179]	Nano-ZnO	EC50	<i>Porcellionides pruinosus</i>	119,000.00	336	10	10	1,190.00
Rousk [180]	Nano-ZnO	EC50	soil bacteria	5,184,000.00	5	10	10	51,840.00
Rousk [180]	Nano-ZnO	EC50	soil bacteria	12,025,000.00	5	10	10	120,250.00

Table S3: Data for sediment toxicity for nano-Ag, nano-TiO<sub>2</sub>, nano-ZnO, CNT and Fullerenes.

Author	ENM	Dose descriptor	Test organism	Concentration (µg/kg)	Exposure time (h)	AF time	AF no-effect	Species sensitivity (µg/kg)
Petersen, et.al. [181]	CNT	HONEC	<i>Lumbriculus variegatus</i> , earthworm	30,000.00	672	1	1	30,000.00
Petersen, et.al. [181]	CNT	HONEC	<i>Lumbriculus variegatus</i>	370,000.00	672	1	1	370,000.00
Kennedy et al. [5]	CNT	LC50	<i>Hyalella azteca</i> , amphipod	264,000,000.00	240	10	10	2,640,000.00
Kennedy et al. [182]	CNT	HONEC	<i>Hyalella azteca</i> , amphipod	99,000,000.00	240	10	1	9,900,000.00
Pakarinen [183]	Fullerenes	HONEC	<i>Lumbriculus variegatus</i>	50,000.00	672	1	1	50,000.00
Musee, et. Al. [184]	Nano-TiO <sub>2</sub>	HONEC	<i>Physa acuta</i> , snail	500,000.00	672	1	1	500,000.00

## References

1. Mouchet, F., et al., *Assessment of the potential in vivo ecotoxicity of double-walled carbon nanotubes (DWNTs) in water, using the amphibian *Amystoma mexicanum**. *Nanotoxicology*, 2007. **1**(2): p. 149-156.
2. Templeton, R.C., et al., *Life-cycle effects of single-walled carbon nanotubes (SWNTs) on an estuarine meiobenthic copepod*. *Environ. Sci. Technol.*, 2006. **40**: p. 7387-7393.
3. Li, M. and C.P. Huang, *The responses of *Ceriodaphnia dubia* toward multi-walled carbon nanotubes: Effect of physical-chemical treatment*. *Carbon*, 2011. **49**(5): p. 1672-1679.
4. Alloy, M.M. and A.P. Roberts, *Effects of suspended multi-walled carbon nanotubes on daphnid growth and reproduction*. *Ecotoxicol Environ Saf*, 2011. **74**(7): p. 1839-43.
5. Kennedy, A.J., et al., *Factors influencing the partitioning and toxicity of nanotubes in the aquatic environment*. *Environmental Toxicology and Chemistry*, 2008. **27**(9): p. 1932-1941.
6. Long, Z.F., et al., *Systematic and Quantitative Investigation of the Mechanism of Carbon Nanotubes' Toxicity toward Algae*. *Environmental Science & Technology*, 2012. **46**(15): p. 8458-8466.
7. Schwab, F., et al., *Are carbon nanotube effects on green algae caused by shading and agglomeration?*. *Environ. Sci. Technol.*, 2011. **45**: p. 6136-6144.
8. Asharani, P.V., et al., *Impact of Multi-Walled Carbon Nanotubes on Aquatic Species*. *Journal of Nanoscience and Nanotechnology*, 2008. **8**(7): p. 3603-3609.
9. Cheng, J., E. Flahaut, and S.H. Cheng, *Effect of carbon nanotubes on developing zebrafish (*Danio rerio*) embryos*. *Environ Toxicol Chem*, 2007. **26**(4): p. 708-16.
10. Zhu, X.S., et al., *Acute toxicities of six manufactured nanomaterial suspensions to *Daphnia magna**. *Journal of Nanoparticle Research*, 2009. **11**(1): p. 67-75.
11. Edgington, A.J., et al., *The influence of natural organic matter on the toxicity of multiwalled carbon nanotubes*. *Environ Toxicol Chem*, 2010. **29**(11): p. 2511-8.
12. Roberts, A.P., et al., *In vivo biomodification of lipid-coated carbon nanotubes by *Daphnia magna**. *Environ. Sci. Technol.*, 2007. **41**: p. 3025-3029.
13. Bayat, N., et al., *The effects of engineered nanoparticles on the cellular structure and growth of *Saccharomyces cerevisiae**. *Nanotoxicology*, 2014. **8**(4): p. 363-73.
14. Zhu, Y., et al., *The Interaction and Toxicity of Multi-Walled Carbon Nanotubes with *Stylynychia Mytilus**. *Journal of Nanoscience and Nanotechnology*, 2006. **6**(5): p. 1357-1364.
15. Ghafari, P., et al., *Impact of carbon nanotubes on the ingestion and digestion of bacteria by ciliated protozoa*. *Nat Nanotechnol*, 2008. **3**(6): p. 347-51.
16. Mouchet, F., et al., *Characterisation and in vivo ecotoxicity evaluation of double-wall carbon nanotubes in larvae of the amphibian *Xenopus laevis**. *Aquat Toxicol*, 2008. **87**(2): p. 127-37.
17. Mouchet, F., et al., *Carbon nanotube ecotoxicity in amphibians: assessment of multiwalled carbon nanotubes and comparison with double-walled carbon nanotubes*. *Nanomedicine (Lond)*, 2010. **5**(6): p. 963-74.
18. Fang, J., et al., *Effect of a fullerene water suspension on bacterial phospholipids and membrane phase behavior*. *Environ. Sci. Technol.*, 2007. **41**: p. 2636-2642.
19. Zhu, X.S., et al., *Oxidative stress and growth inhibition in the freshwater fish *Carassius auratus* induced by chronic exposure to sublethal fullerene aggregates*. *Environmental Toxicology and Chemistry*, 2008. **27**(9): p. 1979-1985.
20. Luo, J., *Toxicity and Bioaccumulation of Nanomaterial in Aquatic Species*. *Journal of the U.S. SJWP*, 2007. **2**.
21. Isaacson, C.W., et al., *Quantification of fullerenes by LC/ESI-MS and its application to in vivo toxicity assays*. *Analytical Chemistry*, 2007. **79**(23): p. 9091-9097.
22. Usenko, C.Y., S.L. Harper, and R.L. Tanguay, *Fullerene C60 exposure elicits an oxidative stress response in embryonic zebrafish*. *Toxicol Appl Pharmacol*, 2008. **229**(1): p. 44-55.
23. Seda, B.C., et al., *Toxicity of aqueous C70-gallic acid suspension in *Daphnia magna**. *Environ Toxicol Chem*, 2012. **31**(1): p. 215-20.
24. Tao, X., et al., *Effects of aqueous stable fullerene nanocrystals (nC60) on *Daphnia magna*: evaluation of sub-lethal reproductive responses and accumulation*. *Chemosphere*, 2009. **77**(11): p. 1482-7.
25. Lovern, S.B. and R. Klaper, **Daphnia magna* mortality when exposed to titanium dioxide and fullerene (C60) nanoparticles*. *Environ. Toxicol. Chem.*, 2006. **25**(4): p. 1132-1137.
26. Zhu, S., E. Oberdorster, and M.L. Haasch, *Toxicity of an engineered nanoparticle (fullerene, C60) in two aquatic species, *Daphnia* and fathead minnow*. *Mar Environ Res*, 2006. **62 Suppl**: p. S5-9.

27. Zhu, X., et al., *Acute toxicities of six manufactured nanomaterial suspensions to Daphnia magna*. Journal of Nanoparticle Research, 2008. **11**(1): p. 67-75.
28. Oberdörster, E., et al., *Ecotoxicology of carbon-based engineered nanoparticles: Effects of fullerene (C<sub>60</sub>) on aquatic organisms*. Carbon, 2006. **44**: p. 1112-1120.
29. Ivask, A., et al., *Profiling of the reactive oxygen species-related ecotoxicity of CuO, ZnO, TiO<sub>2</sub>, silver and fullerene nanoparticles using a set of recombinant luminescent Escherichia coli strains: differentiating the impact of particles and solubilised metals*. Anal Bioanal Chem, 2010. **398**(2): p. 701-16.
30. Oberdörster, E., *Manufactured nanomaterials (Fullerenes, C<sub>60</sub>) induce oxidative stress in the brain of juvenile largemouth bass*. Environ. Health Perspect., 2004. **112**(10): p. 1058-1062.
31. Gao, J., et al., *Effects of engineered nanomaterials on microbial catalyzed biogeochemical processes in sediments*. J Hazard Mater, 2011. **186**(1): p. 940-5.
32. Luo, Z., et al., *Gold and silver nanoparticle effects on ammonia-oxidizing bacteria cultures under ammonoxidation*. Chemosphere, 2014.
33. Jin, X., et al., *High-Throughput Screening of Silver Nanoparticle Stability and Bacterial Inactivation in Aquatic Media: Influence of Specific Ions*. Environmental Science & Technology, 2010. **44**(19): p. 7321-7328.
34. Yoon, K.Y., et al., *Susceptibility constants of Escherichia coli and Bacillus subtilis to silver and copper nanoparticles*. Sci Total Environ, 2007. **373**(2-3): p. 572-5.
35. Roh, J.Y., et al., *Ecotoxicity of Silver Nanoparticles on the Soil Nematode Caenorhabditis elegans Using Functional Ecotoxicogenomics*. Environmental Science & Technology, 2009. **43**(10): p. 3933-3940.
36. Kim, S.W., S.H. Nam, and Y.J. An, *Interaction of silver nanoparticles with biological surfaces of Caenorhabditis elegans*. Ecotoxicol Environ Saf, 2012. **77**: p. 64-70.
37. Meyer, J.N., et al., *Intracellular uptake and associated toxicity of silver nanoparticles in Caenorhabditis elegans*. Aquatic Toxicology, 2010. **100**(2): p. 140-150.
38. Angel, B.M., et al., *The impact of size on the fate and toxicity of nanoparticulate silver in aquatic systems*. Chemosphere, 2013. **93**(2): p. 359-65.
39. Gao, J., et al., *Dispersion and toxicity of selected manufactured nanomaterials in natural river water samples: Effects of water chemical composition*. Environ. Sci. Technol., 2009. **43**: p. 3322-3328.
40. Kennedy, A.J., et al., *Impact of Organic Carbon on the Stability and Toxicity of Fresh and Stored Silver Nanoparticles*. Environmental Science & Technology, 2012. **46**(19): p. 10772-10780.
41. Griffith, R.J., et al., *Effects of particle composition and species on toxicity of metallic nanomaterials in aquatic organisms*. Environmental Toxicology and Chemistry, 2008. **27**(9): p. 1972-1978.
42. McLaughlin, J. and J.C.J. Bonzongo, *Effects of natural water chemistry on nanosilver behavior and toxicity to Ceriodaphnia dubia and Pseudokirchneriella subcapitata*. Environmental Toxicology and Chemistry, 2012. **31**(1): p. 168-175.
43. Nair, P.M., S.Y. Park, and J. Choi, *Evaluation of the effect of silver nanoparticles and silver ions using stress responsive gene expression in Chironomus riparius*. Chemosphere, 2013. **92**(5): p. 592-9.
44. Navarro, E., et al., *Toxicity of Silver Nanoparticles to Chlamydomonas reinhardtii*. Environ. Sci. Technol., 2008. **42**(23): p. 8959-8964.
45. Wang, Z., et al., *Aquatic toxicity of nanosilver colloids to different trophic organisms: contributions of particles and free silver ion*. Environ Toxicol Chem, 2012. **31**(10): p. 2408-13.
46. Massarsky, A., et al., *Assessment of nanosilver toxicity during zebrafish (Danio rerio) development*. Chemosphere, 2013. **92**(1): p. 59-66.
47. Kaewamatawong, T., Ponpornpisit, A., et al., *Toxicity Test of Nanosilver Particles on Zebrafish (Danio rerio) Embryonic Development*. Thai J Vet Med., 2012. **42**(3): p. 305-310.
48. Cunningham, S., et al., *Effect of nanoparticle stabilization and physicochemical properties on exposure outcome: acute toxicity of silver nanoparticle preparations in zebrafish (Danio rerio)*. Environ Sci Technol, 2013. **47**(8): p. 3883-92.
49. Bilberg, K., et al., *In Vivo Toxicity of Silver Nanoparticles and Silver Ions in Zebrafish (Danio rerio)*. J Toxicol, 2012. **2012**: p. 293784.
50. Muth-Kohne, E., et al., *The toxicity of silver nanoparticles to zebrafish embryos increases through sewage treatment processes*. Ecotoxicology, 2013. **22**(8): p. 1264-77.
51. van Aerle, R., et al., *Molecular mechanisms of toxicity of silver nanoparticles in zebrafish embryos*. Environ Sci Technol, 2013. **47**(14): p. 8005-14.
52. Asharani, P.V., et al., *Toxicity of silver nanoparticles in zebrafish models*. Nanotechnology, 2008. **19**(25): p. 255102.
53. Choi, J.E., et al., *Induction of oxidative stress and apoptosis by silver nanoparticles in the liver of adult zebrafish*. Aquat Toxicol, 2010. **100**(2): p. 151-9.

54. Volker, C., et al., *Comparative toxicity assessment of nanosilver on three Daphnia species in acute, chronic and multi-generation experiments*. PLoS One, 2013. **8**(10): p. e75026.
55. Lee, Y.J., et al., *Ion-release kinetics and ecotoxicity effects of silver nanoparticles*. Environmental Toxicology and Chemistry, 2012. **31**(1): p. 155-159.
56. Kim, J., S. Kim, and S. Lee, *Differentiation of the toxicities of silver nanoparticles and silver ions to the Japanese medaka (Oryzias latipes) and the cladoceran Daphnia magna*. Nanotoxicology, 2011. **5**(2): p. 208-214.
57. Zhao, C.M. and W.X. Wang, *Importance of surface coatings and soluble silver in silver nanoparticles toxicity to Daphnia magna*. Nanotoxicology, 2012. **6**(4): p. 361-70.
58. Allen, H.J., et al., *Effects from filtration, capping agents, and presence/absence of food on the toxicity of silver nanoparticles to Daphnia magna*. Environ Toxicol Chem, 2010. **29**(12): p. 2742-50.
59. Georgantzopoulou, A., et al., *Ag nanoparticles: size- and surface-dependent effects on model aquatic organisms and uptake evaluation with NanoSIMS*. Nanotoxicology, 2013. **7**: p. 1168-78.
60. Poynton, H.C., et al., *Toxicogenomic Responses of Nanotoxicity in Daphnia magna Exposed to Silver Nitrate and Coated Silver Nanoparticles*. Environmental Science & Technology, 2012. **46**(11): p. 6288-6296.
61. Kennedy, A.J., et al., *Fractionating Nanosilver: Importance for Determining Toxicity to Aquatic Test Organisms*. Environmental Science & Technology, 2010. **44**(24): p. 9571-9577.
62. Asghari, S., et al., *Toxicity of various silver nanoparticles compared to silver ions in Daphnia magna*. J Nanobiotechnology, 2012. **10**: p. 14.
63. Newton, K.M., et al., *Silver nanoparticle toxicity to Daphnia magna is a function of dissolved silver concentration*. Environmental Toxicology and Chemistry, 2013. **32**(10): p. 2356-2364.
64. Das, P., M.A. Xenopoulos, and C.D. Metcalfe, *Toxicity of silver and titanium dioxide nanoparticle suspensions to the aquatic invertebrate, Daphnia magna*. Bull Environ Contam Toxicol, 2013. **91**(1): p. 76-82.
65. Li, T., et al., *Comparative toxicity study of Ag, Au, and Ag-Au bimetallic nanoparticles on Daphnia magna*. Analytical and Bioanalytical Chemistry, 2010. **398**(2): p. 689-700.
66. Jo, H.J., et al., *Acute toxicity of Ag and CuO nanoparticle suspensions against Daphnia magna: the importance of their dissolved fraction varying with preparation methods*. J Hazard Mater, 2012. **227-228**: p. 301-8.
67. Hoheisel, S.M., S. Diamond, and D. Mount, *Comparison of nanosilver and ionic silver toxicity in Daphnia magna and Pimephales promelas*. Environmental Toxicology and Chemistry, 2012. **31**(11): p. 2557-2563.
68. Stensberg, M.C., et al., *Silver nanoparticle-specific mitotoxicity in Daphnia magna*. Nanotoxicology, 2014. **8**: p. 833-42.
69. Zhao, C.M. and W.X. Wang, *Comparison of acute and chronic toxicity of silver nanoparticles and silver nitrate to Daphnia magna*. Environ Toxicol Chem, 2011. **30**(4): p. 885-92.
70. Pokhrel, L.R., B. Dubey, and P.R. Scheuerman, *Impacts of select organic ligands on the colloidal stability, dissolution dynamics, and toxicity of silver nanoparticles*. Environ Sci Technol, 2013. **47**(22): p. 12877-85.
71. Blinova, I., et al., *Toxicity of two types of silver nanoparticles to aquatic crustaceans Daphnia magna and Thamnocephalus platyurus*. Environ Sci Pollut Res Int, 2013. **20**(5): p. 3456-63.
72. Martínez-Castañón, G.A., et al., *Synthesis and antibacterial activity of silver nanoparticles with different sizes*. Journal of Nanoparticle Research, 2008. **10**(8): p. 1343-1348.
73. Pokhrel, L.R., et al., *Rapid screening of aquatic toxicity of several metal-based nanoparticles using the MetPLATE bioassay*. Sci Total Environ, 2012. **426**: p. 414-22.
74. Arce Funck, J., et al., *Behavioural and physiological responses of Gammarus fossarum (Crustacea Amphipoda) exposed to silver*. Aquat Toxicol, 2013. **142-143**: p. 73-84.
75. Gubbins, E.J., L.C. Batty, and J.R. Lead, *Phytotoxicity of silver nanoparticles to Lemna minor L.* Environ Pollut, 2011. **159**(6): p. 1551-9.
76. Kim, E., et al., *Growth inhibition of aquatic plant caused by silver and titanium oxide nanoparticles*. Toxicology and Environmental Health Sciences, 2011. **3**(1): p. 1-6.
77. Ali, D., et al., *Sensitivity of freshwater pulmonate snail Lymnaea luteola L., to silver nanoparticles*. Chemosphere, 2014. **104**: p. 134-40.
78. Miao, A.J., et al., *Intracellular Uptake: A Possible Mechanism for Silver Engineered Nanoparticle Toxicity to a Freshwater Alga Ochromonas danica*. Plos One, 2010. **5**(12).
79. Govindasamy, R. and A.A. Rahuman, *Histopathological studies and oxidative stress of synthesized silver nanoparticles in Mozambique tilapia (Oreochromis mossambicus)*. Journal of Environmental Sciences, 2012. **24**(6): p. 1091-1098.
80. Chae, Y.J., et al., *Evaluation of the toxic impact of silver nanoparticles on Japanese medaka (Oryzias latipes)*. Aquat Toxicol, 2009. **94**(4): p. 320-7.
81. Wu, Y., et al., *Effects of silver nanoparticles on the development and histopathology biomarkers of Japanese medaka (Oryzias latipes) using the partial-life test*. Aquat Toxicol, 2010. **100**(2): p. 160-7.



82. Kashiwada, S., et al., *Silver nanocolloids disrupt medaka embryogenesis through vital gene expressions*. Environ Sci Technol, 2012. **46**(11): p. 6278-87.
83. Kim, J.Y., et al., *Developmental toxicity of Japanese medaka embryos by silver nanoparticles and released ions in the presence of humic acid*. Ecotoxicol Environ Saf, 2013. **92**: p. 57-63.
84. Kvittek, L., et al., *Initial Study on the Toxicity of Silver Nanoparticles (NPs) against Paramecium caudatum*. Journal of Physical Chemistry C, 2009. **113**(11): p. 4296-4300.
85. Bilberg, K., et al., *Silver nanoparticles and silver nitrate cause respiratory stress in Eurasian perch (Perca fluviatilis)*. Aquat Toxicol, 2010. **96**(2): p. 159-65.
86. Laban, G., et al., *The effects of silver nanoparticles on fathead minnow (Pimephales promelas) embryos*. Ecotoxicology, 2010. **19**(1): p. 185-195.
87. Debabrata D., G.A., *Cellular responses of Saccharomyces cerevisiae to silver nanoparticles*. Res J Biotech, 2013. **8**(1): p. 11.
88. Niazi, J.H., et al., *Global gene response in Saccharomyces cerevisiae exposed to silver nanoparticles*. Appl Biochem Biotechnol, 2011. **164**(8): p. 1278-91.
89. Jiang, H.S., et al., *Physiological analysis of silver nanoparticles and AgNO<sub>3</sub> toxicity to Spirodela polyrhiza*. Environ Toxicol Chem, 2012. **31**(8): p. 1880-6.
90. Burchardt, A.D., et al., *Effects of silver nanoparticles in diatom Thalassiosira pseudonana and cyanobacterium Synechococcus sp.* Environ Sci Technol, 2012. **46**(20): p. 11336-44.
91. Juganson, K., et al., *Extracellular conversion of silver ions into silver nanoparticles by protozoan Tetrahymena thermophila*. Environmental Science: Processes & Impacts, 2013. **15**(1): p. 244-250.
92. Cherchi, C., et al., *Impact of nano titanium dioxide exposure on cellular structure of Anabaena variabilis and evidence of internalization*. Environ Toxicol Chem, 2011. **30**(4): p. 861-9.
93. Adams, L.K., D.Y. Lyon, and P.J. Alvarez, *Comparative eco-toxicity of nanoscale TiO<sub>2</sub>, SiO<sub>2</sub>, and ZnO water suspensions*. Water Res, 2006. **40**(19): p. 3527-32.
94. Clement, L., C. Hurel, and N. Marmier, *Toxicity of TiO<sub>2</sub> nanoparticles to cladocerans, algae, rotifers and plants - effects of size and crystalline structure*. Chemosphere, 2013. **90**(3): p. 1083-90.
95. Wu, Q., et al., *Comparison of toxicities from three metal oxide nanoparticles at environmental relevant concentrations in nematode Caenorhabditis elegans*. Chemosphere, 2013. **90**(3): p. 1123-31.
96. Wang, H., R.L. Wick, and B. Xing, *Toxicity of nanoparticulate and bulk ZnO, Al<sub>2</sub>O<sub>3</sub> and TiO<sub>2</sub> to the nematode Caenorhabditis elegans*. Environ Pollut, 2009. **157**(4): p. 1171-7.
97. Hall, S., et al., *Acute and chronic toxicity of nano-scale TiO<sub>2</sub> particles to freshwater fish, cladocerans, and green algae, and effects of organic and inorganic substrate on TiO<sub>2</sub> toxicity*. Nanotoxicology, 2009. **3**(2): p. 91-97.
98. Wang, D., et al., *Synergistic toxic effect of nano-TiO and As(V) on Ceriodaphnia dubia*. Sci Total Environ, 2011. **409**(7): p. 1351-6.
99. Li, S., et al., *Species sensitivity and dependence on exposure conditions impacting the phototoxicity of TiO<sub>2</sub> nanoparticles to benthic organisms*. Environ Toxicol Chem, 2014. **33**(7): p. 1563-9.
100. Wang, J., et al., *Toxicity assessment of manufactured nanomaterials using the unicellular green alga Chlamydomonas reinhardtii*. Chemosphere, 2008. **73**(7): p. 1121-8.
101. Gunawan, C., et al., *Submicron and nano formulations of titanium dioxide and zinc oxide stimulate unique cellular toxicological responses in the green microalga Chlamydomonas reinhardtii*. J Hazard Mater, 2013. **260**: p. 984-92.
102. Sadiq, I.M., et al., *Ecotoxicity study of titania (TiO<sub>2</sub>) NPs on two microalgae species: Scenedesmus sp. and Chlorella sp.* Ecotoxicol Environ Saf, 2011. **74**(5): p. 1180-7.
103. Ji, J., Z.F. Long, and D.H. Lin, *Toxicity of oxide nanoparticles to the green algae Chlorella sp.* Chemical Engineering Journal, 2011. **170**(2-3): p. 525-530.
104. Chen, J., et al., *Effects of titanium dioxide nano-particles on growth and some histological parameters of zebrafish (Danio rerio) after a long-term exposure*. Aquat Toxicol, 2011. **101**(3-4): p. 493-9.
105. Xiong, D., et al., *Effects of nano-scale TiO<sub>2</sub>, ZnO and their bulk counterparts on zebrafish: acute toxicity, oxidative stress and oxidative damage*. Sci Total Environ, 2011. **409**(8): p. 1444-52.
106. Yang, S.P., et al., *Influence of humic acid on titanium dioxide nanoparticle toxicity to developing zebrafish*. Environ Sci Technol, 2013. **47**(9): p. 4718-25.
107. Bar-Ilan, O., et al., *Titanium dioxide nanoparticles produce phototoxicity in the developing zebrafish*. Nanotoxicology, 2012. **6**(6): p. 670-9.
108. Zhu, X., et al., *Comparative toxicity of several metal oxide nanoparticle aqueous suspensions to Zebrafish (Danio rerio) early developmental stage*. J Environ Sci Health A Tox Hazard Subst Environ Eng, 2008. **43**(3): p. 278-84.
109. Ma, H., A. Brennan, and S.A. Diamond, *Phototoxicity of TiO<sub>2</sub> nanoparticles under solar radiation to two aquatic species: Daphnia magna and Japanese medaka*. Environmental Toxicology Chemistry, 2012. **31**(7): p. 1621-1629.

110. Dabrunz, A., et al., *Biological Surface Coating and Molting Inhibition as Mechanisms of TiO<sub>2</sub> Nanoparticle Toxicity in Daphnia magna*. Plos One, 2011. **6**(5).
111. Zhu, X., Y. Chang, and Y. Chen, *Toxicity and bioaccumulation of TiO<sub>2</sub> nanoparticle aggregates in Daphnia magna*. Chemosphere, 2010. **78**(3): p. 209-15.
112. Warheit, D.B., et al., *Development of a base set of toxicity tests using ultrafine TiO<sub>2</sub> particles as a component of nanoparticle risk management*. Toxicol Lett, 2007. **171**(3): p. 99-110.
113. Kim, K.T., et al., *Oxidative stress responses of Daphnia magna exposed to TiO<sub>2</sub> nanoparticles according to size fraction*. Science of the Total Environment, 2010. **408**(10): p. 2268-2272.
114. Jacobasch, C., et al., *Long-term effects of nanoscaled titanium dioxide on the cladoceran Daphnia magna over six generations*. Environ Pollut, 2013. **186C**: p. 180-186.
115. Amiano, I., et al., *Acute toxicity of nanosized TiO<sub>2</sub> to Daphnia magna under UVA irradiation*. Environ Toxicol Chem, 2012. **31**(11): p. 2564-6.
116. Wiench, K., et al., *Acute and chronic effects of nano- and non-nano-scale TiO<sub>2</sub> and ZnO particles on mobility and reproduction of the freshwater invertebrate Daphnia magna*. Chemosphere, 2009. **76**(10): p. 1356-65.
117. Heinlaan, M., et al., *Toxicity of nanosized and bulk ZnO, CuO and TiO<sub>2</sub> to bacteria Vibrio fischeri and crustaceans Daphnia magna and Thamnocephalus platyurus*. Chemosphere, 2008. **71**(7): p. 1308-1316.
118. Marcone, G.P.S., et al., *Ecotoxicity of TiO<sub>2</sub> to Daphnia similis under irradiation*. Journal of Hazardous Materials, 2012. **211**: p. 436-442.
119. Hund-Rinke, K. and M. Simon, *Ecotoxic effect of photocatalytic active nanoparticles TiO<sub>2</sub> on algae and daphnids*. Environ. Sci. Pollution Res., 2006. **13**(4): p. 225-232.
120. Bigorgne, E., et al., *Ecotoxicological assessment of TiO<sub>2</sub> byproducts on the earthworm Eisenia fetida*. Environ Pollut, 2011. **159**(10): p. 2698-705.
121. Dasari, T.P., K. Pathakoti, and H.-M. Hwang, *Determination of the mechanism of photoinduced toxicity of selected metal oxide nanoparticles (ZnO, CuO, Co<sub>3</sub>O<sub>4</sub> and TiO<sub>2</sub>) to E. coli bacteria*. Journal of Environmental Sciences, 2013. **25**(5): p. 882-888.
122. Hu, X., et al., *In vitro evaluation of cytotoxicity of engineered metal oxide nanoparticles*. Sci Total Environ, 2009. **407**(8): p. 3070-2.
123. Bundschuh, M., et al., *Effects of nano-TiO<sub>2</sub> in combination with ambient UV-irradiation on a leaf shredding amphipod*. Chemosphere, 2011. **85**(10): p. 1563-7.
124. Zhu, X., J. Zhou, and Z. Cai, *TiO<sub>2</sub> nanoparticles in the marine environment: impact on the toxicity of tributyltin to abalone (Haliotis diversicolor supertexta) embryos*. Environ Sci Technol, 2011. **45**(8): p. 3753-8.
125. Li, S., et al., *Phototoxicity of TiO<sub>2</sub> nanoparticles to a freshwater benthic amphipod: are benthic systems at risk?* Sci Total Environ, 2014. **466-467**: p. 800-8.
126. Federici, G., B.J. Shaw, and R.D. Handy, *Toxicity of titanium dioxide nanoparticles to rainbow trout (Oncorhynchus mykiss): gill injury, oxidative stress, and other physiological effects*. Aquat Toxicol, 2007. **84**(4): p. 415-30.
127. Li, K., et al., *Surface interactions affect the toxicity of engineered metal oxide nanoparticles toward Paramecium*. Chem Res Toxicol, 2012. **25**(8): p. 1675-81.
128. Jovanovic, B., et al., *Effects of nanosized titanium dioxide on innate immune system of fathead minnow (Pimephales promelas Rafinesque, 1820)*. Ecotoxicology and Environmental Safety, 2011. **74**(4): p. 675-683.
129. Kim, S., J. Kim, and I. Lee, *Effects of Zn and ZnO nanoparticles and Zn<sup>2+</sup> on soil enzyme activity and bioaccumulation of Zn in Cucumis sativus*. Chemistry and Ecology, 2011. **27**(1): p. 49-55.
130. Aruoja, V., et al., *Toxicity of nanoparticles of CuO, ZnO and TiO<sub>2</sub> to microalgae Pseudokirchneriella subcapitata*. Sci Total Environ, 2009. **407**(4): p. 1461-8.
131. Hartmann, N.B., et al., *Algal testing of titanium dioxide nanoparticles-Testing considerations, inhibitory effects and modification of cadmium bioavailability*. Toxicology, 2010. **269**(2-3): p. 190-197.
132. Metzler, D.M., et al., *Responses of algae to photocatalytic nano-TiO<sub>2</sub> particles with an emphasis on the effect of particle size*. Chemical Engineering Journal, 2011. **170**(2-3): p. 538-546.
133. Kasemets, K., et al., *Toxicity of nanoparticles of ZnO, CuO and TiO<sub>2</sub> to yeast Saccharomyces cerevisiae*. Toxicol In Vitro, 2009. **23**(6): p. 1116-22.
134. Velzeboer, I., et al., *Aquatic ecotoxicity tests of some nanomaterials*. Environmental Toxicology and Chemistry, 2008. **27**(9): p. 1942-1947.
135. Nations, S., et al., *Acute effects of Fe<sub>2</sub>O<sub>3</sub>, TiO<sub>2</sub>, ZnO and CuO nanomaterials on Xenopus laevis*. Chemosphere, 2011. **83**(8): p. 1053-61.
136. Li, M., et al., *Stability, Bioavailability, and Bacterial Toxicity of ZnO and Iron-Doped ZnO Nanoparticles in Aquatic Media*. Environmental Science & Technology, 2010. **45**(2): p. 755-761.

137. Jones, N., et al., *Antibacterial activity of ZnO nanoparticle suspensions on a broad spectrum of microorganisms*. FEMS Microbiol Lett, 2008. **279**(1): p. 71-6.
138. Ma, H., Besrtsch, P., Glenn, T. T., Kabengl, N., Williams, P., *Toxicity of manufactured zinc oxide nanoparticles in the nematode Caenorhabditis elegans*. Environmental Toxicology and Chemistry, 2009. **28**(6): p. 1324–1330.
139. Ma, H., et al., *Comparative phototoxicity of nanoparticulate and bulk ZnO to a free-living nematode Caenorhabditis elegans: The importance of illumination mode and primary particle size*. Environmental Pollution, 2011. **159**(6): p. 1473-1480.
140. Lin, D.H., et al., *The influence of dissolved and surface-bound humic acid on the toxicity of TiO<sub>2</sub> nanoparticles to Chlorella sp.* Water Research, 2012. **46**(14): p. 4477-4487.
141. Fabrega, J., et al., *Sequestration of zinc from zinc oxide nanoparticles and life cycle effects in the sediment dweller amphipod Corophium volutator*. Environ Sci Technol, 2012. **46**(2): p. 1128-35.
142. Yu, L.P., et al., *Comparative toxicity of nano-ZnO and bulk ZnO suspensions to zebrafish and the effects of sedimentation, OH production and particle dissolution in distilled water*. J Environ Monit, 2011. **13**(7): p. 1975-82.
143. Zhao, H., Lu, G., Jin, S., *Toxicity of Nanoscale CuO and ZnO to Daphnia magna*. CHEM. RES. CHINESE UNIVERSITIES 2012. **28**(2): p. 209—213.
144. Lopes, S., et al., *Zinc oxide nanoparticles toxicity to Daphnia magna: size-dependent effects and dissolution*. Environ Toxicol Chem, 2014. **33**(1): p. 190-8.
145. Blinova, I., et al., *Ecotoxicity of nanoparticles of CuO and ZnO in natural water*. Environ Pollut, 2010. **158**(1): p. 41-7.
146. Naddafi, K., M.R. Zare, and S. Nazmara, *Investigating potential toxicity of phenanthrene adsorbed to nano-ZnO using Daphnia magna*. Toxicological & Environmental Chemistry, 2011. **93**(4): p. 729-737.
147. Poynton, H.C., et al., *Differential Gene Expression in Daphnia magna Suggests Distinct Modes of Action and Bioavailability for ZnO Nanoparticles and Zn Ions*. Environmental Science & Technology, 2011. **45**(2): p. 762-768.
148. Zarrindokht, E.-K., *Antibacterial activity of ZnO nanoparticle on Gram-positive and Gram-negative bacteria*. African Journal of Microbiology Research, 2012. **5**(18).
149. Poynton, H.C., et al., *Toxicity and transcriptomic analysis in Hyalella azteca suggests increased exposure and susceptibility of epibenthic organisms to zinc oxide nanoparticles*. Environ Sci Technol, 2013. **47**(16): p. 9453-60.
150. Miller, R.J., et al., *Impacts of Metal Oxide Nanoparticles on Marine Phytoplankton*. Environmental Science & Technology, 2010. **44**(19): p. 7329-7334.
151. Lin, D. and B. Xing, *Phytotoxicity of nanoparticles: Inhibition of seed germination and root growth*. Environ. Pollut., 2007. **150**: p. 243-250.
152. Lin, D. and B. Xing, *Root uptake and phytotoxicity of ZnO nanoparticles*. Environ. Sci. Technol., 2008. **42**(15): p. 5580-5585.
153. Franklin, N.M., et al., *Comparative toxicity of nanoparticulate ZnO, bulk ZnO, and ZnCl<sub>2</sub> to a freshwater microalga (Pseudokirchneriella subcapitata): The importance of particle solubility*. Environmental Science & Technology, 2007. **41**(24): p. 8484-8490.
154. Lee, W.M. and Y.J. An, *Effects of zinc oxide and titanium dioxide nanoparticles on green algae under visible, UVA, and UVB irradiations: no evidence of enhanced algal toxicity under UV pre-irradiation*. Chemosphere, 2013. **91**(4): p. 536-44.
155. Mortimer, M., K. Kasemets, and A. Kahru, *Toxicity of ZnO and CuO nanoparticles to ciliated protozoa Tetrahymena thermophila*. Toxicology, 2010. **269**(2-3): p. 182-9.
156. Wong, S.W., et al., *Toxicities of nano zinc oxide to five marine organisms: influences of aggregate size and ion solubility*. Anal Bioanal Chem, 2010. **396**(2): p. 609-18.
157. Jarvis, T.A., et al., *Toxicity of ZnO nanoparticles to the copepod Acartia tonsa, exposed through a phytoplankton diet*. Environmental Toxicology and Chemistry, 2013. **32**(6): p. 1264-1269.
158. De La Torre-Roche, R., et al., *Multiwalled carbon nanotubes and c60 fullerenes differentially impact the accumulation of weathered pesticides in four agricultural plants*. Environ Sci Technol, 2013. **47**(21): p. 12539-47.
159. Scott-Fordsmand, J.J., et al., *The toxicity testing of double-walled nanotubes-contaminated food to Eisenia veneta earthworms*. Ecotoxicol Environ Saf, 2008. **71**(3): p. 616-9.
160. Chung, H., et al., *The effect of multi-walled carbon nanotubes on soil microbial activity*. Ecotoxicol Environ Saf, 2011. **74**(4): p. 569-75.
161. Shrestha, B., et al., *An evaluation of the impact of multiwalled carbon nanotubes on soil microbial community structure and functioning*. J Hazard Mater, 2013. **261**: p. 188-97.
162. Johansen, A., et al., *Effects of C-60 fullerene nanoparticles on soil bacteria and protozoans*. Environmental Toxicology and Chemistry, 2008. **27**(9): p. 1895-1903.
163. van der Ploeg, M.J., et al., *Effects of C60 nanoparticle exposure on earthworms (Lumbricus rubellus) and implications for population dynamics*. Environ Pollut, 2011. **159**(1): p. 198-203.
164. Tong, Z., et al., *Impact of fullerene (C<sub>60</sub>) on a soil microbial community*. Environ. Sci. Technol., 2007. **41**: p. 2985-2991.

165. Schlich, K., et al., *Effects of silver nanoparticles and silver nitrate in the earthworm reproduction test*. Environ Toxicol Chem, 2013. **32**(1): p. 181-8.
166. Shoultz-Wilson, W.A., et al., *Role of Particle Size and Soil Type in Toxicity of Silver Nanoparticles to Earthworms*. Soil Science Society of America Journal, 2011. **75**(2): p. 365.
167. Gomes, S.I., et al., *Mechanisms of response to silver nanoparticles on Enchytraeus albidus (Oligochaeta): survival, reproduction and gene expression profile*. J Hazard Mater, 2013. **254-255**: p. 336-44.
168. Lee, W.M., J.I. Kwak, and Y.J. An, *Effect of silver nanoparticles in crop plants Phaseolus radiatus and Sorghum bicolor: Media effect on phytotoxicity*. Chemosphere, 2012. **86**(5): p. 491-499.
169. Heckmann, L.H., et al., *Limit-test toxicity screening of selected inorganic nanoparticles to the earthworm Eisenia fetida*. Ecotoxicology, 2011. **20**(1): p. 226-233.
170. Hu, C.W., et al., *Toxicological effects of TiO<sub>2</sub> and ZnO nanoparticles in soil on earthworm Eisenia fetida*. Soil Biology & Biochemistry, 2010. **42**(4): p. 586-591.
171. Canas, J.E., et al., *Acute and reproductive toxicity of nano-sized metal oxides (ZnO and TiO<sub>2</sub>) to earthworms (Eisenia fetida)*. J Environ Monit, 2011. **13**(12): p. 3351-7.
172. Nogueira, V., et al., *Impact of organic and inorganic nanomaterials in the soil microbial community structure*. Science of the Total Environment, 2012. **424**: p. 344-350.
173. Heggelund, L.R., et al., *Soil pH effects on the comparative toxicity of dissolved zinc, non-nano and nano ZnO to the earthworm Eisenia fetida*. Nanotoxicology, 2014. **8**(5): p. 559-72.
174. Hooper, H.L., et al., *Comparative chronic toxicity of nanoparticulate and ionic zinc to the earthworm Eisenia veneta in a soil matrix*. Environ Int, 2011. **37**(6): p. 1111-7.
175. Waalewijn-Kool, P.L., et al., *The effect of pH on the toxicity of zinc oxide nanoparticles to Folsomia candida in amended field soil*. Environ Toxicol Chem, 2013. **32**(10): p. 2349-55.
176. Waalewijn-Kool, P.L., et al., *Sorption, dissolution and pH determine the long-term equilibration and toxicity of coated and uncoated ZnO nanoparticles in soil*. Environ Pollut, 2013. **178**: p. 59-64.
177. Kool, P.L., M.D. Ortiz, and C.A.M. van Geste, *Chronic toxicity of ZnO nanoparticles, non-nano ZnO and ZnCl<sub>2</sub> to Folsomia candida (Collembola) in relation to bioavailability in soil*. Environmental Pollution, 2011. **159**(10): p. 2713-2719.
178. Manzo, S., et al., *Investigation of ZnO nanoparticles' ecotoxicological effects towards different soil organisms*. Environmental Science and Pollution Research, 2010.
179. Tourinho, P.S., et al., *Influence of soil pH on the toxicity of zinc oxide nanoparticles to the terrestrial isopod Porcellionides pruinosus*. Environ Toxicol Chem, 2013. **32**(12): p. 2808-15.
180. Rousk, J., et al., *Comparative toxicity of nanoparticulate CuO and ZnO to soil bacterial communities*. PLoS One, 2012. **7**(3): p. e34197.
181. Petersen, E.J., Q. Huang, and W.J. Weber, *Ecological uptake and depuration of carbon nanotubes by Lumbriculus variegatus*. Environ. Health Perspect., 2008. **116**(4): p. 496-500.
182. Kennedy, A.J., et al., *INFLUENCE OF NANOTUBE PREPARATION IN AQUATIC BIOASSAYS*. Environmental Toxicology and Chemistry, 2009. **28**(9): p. 1930-1938.
183. Pakarinen, K., et al., *Adverse effects of fullerenes (nC(60)) spiked to sediments on Lumbriculus variegatus (Oligochaeta)*. Environmental Pollution, 2011. **159**(12): p. 3750-3756.
184. Musee, N., et al., *The effects of engineered nanoparticles on survival, reproduction, and behaviour of freshwater snail, Physa acuta (Draparnaud, 1805)*. Chemosphere, 2010. **81**(10): p. 1196-1203.