Supplementary methods S1:

Detailed RT and PCR conditions:

RT was performed using Superscript II (Invitrogen) using the following protocol: 8µL viral RNA was incubated with 2µL of DMSO for 4 minutes at 94°C then chilled on ice. Five microliters of the DMSO-treated RNA was then added to a mix containing 4µL of dNTPs (GeneAmp dNTP Blend, 2.5mM each, Applied Biosystems), 0.2µL of primer A-NCR32- or B-NCR20- (for segment A and B respectively, at 10µM) and 2.8µL RNase-free water. This mix was heated to 65°C for 5 minutes and then chilled on ice. A mix containing 4µL of first-strand buffer (from the superscript II kit, Invitrogen), 2µL of DTT (0.1M) and 40U of RNasin inhibitor (Promega) was then gently added. The reaction was incubated for 2 minutes at 42°C. Finally, 200U of Superscript II was added to the reaction, which was heated to 42°C for 50 minutes. The enzyme was inactivated by heating the reaction to 70°C for 15 minutes.

PCR was carried out using the Expand High Fidelity kit (Roche). Four microliters of RT product were added to a mix containing 9.7µL RNase-free water, 3.2µL dNTPs (1.25mM), 2µL of MgCl2 (15mM), 0.3µL enzyme and 0.4µL of each primer (pre-diluted to 15µM, respectively A-NCR32- and A-NCR31+ for segment A and B-NCR20- and B-NCR25+ for segment B). PCR program included the following steps: 2min at 94°C, ten cycles comprising 15s at 94°C, 30s at respectively 64.8°C and 60.6°c for segment A and B, 2min 30s at 68°C followed by 25 cycles comprising 15s at 94°C, 30s at respectively 64.8°C and 60.6°c for segment A and B, and 2min 30s at 68°C with an increment of 5s per cycle and a final step at 68°C for 7 min.