**Supporting methods S1**

**16S rDNA amplification**

PCR reactions were carried out in 25 µl volume containing 1X Flexi PCR buffer (Promega, Madison, WI, USA), 1.5 mM MgCl2, 250 µM deoxynucleotide triphosphates (dNTPs), 400 nM of each primer, 1U GoTaq®Flexi DNA polymerase (Promega) and 0.5% tween 20 (v/v). Amplifications were performed using a Tetrad 2 thermal cycler (Bio-Rad) under the following conditions: an initial denaturation for 4 min at 95 °C, 5 cycles of 95 °C for 30 s, 60 °C for 30 s, 72 °C for 45 s, 5 cycles of 95 °C for 30 s, 60 °C for 30 s (decreasing temperature by 1°C at each cycle), 72 °C for 45 s, 25 cycles of 95 °C for 30 s, 55 °C for 30 s, 72 °C for 45 s and a final extension step at 72°C for 10 min. Amplicons were purified with PureLinkTMPCR purification kit (Invitrogen, Thermo Fisher Scientific) and subjected to sequencing at the Interdepartmental Centre for Agricultural, Chemical and Industrial Biotechnology (CIBIACI). The 16S rDNA sequence chromatograms were checked and edited to verify the absence of ambiguous peaks.