

Supplemental Figure 1. Two commercially available $ER\alpha$ antibodies were used to perform western blots on cell lysate from Ishikawa cells. All antibodies show numerous nonspecific bands, precluding their use in assays utilizing immunofluorescent labeling. For each antibody, the targeted receptor epitope is listed.

PROPOSED MODEL FOR ESTROGEN-INITIATED SIGNALING IN THE HUMAN ENDOMETRIUM

We created a mathematical representation of the signaling diagram in Figure 3A assuming Michaelis-Menten binding and mass-action dynamics. We assume that a ligand binds $ER\alpha 66$, $ER\beta$, $ER\alpha 46$, and GPER according to Michaelis-Menten kinetics. The K_m and V_{max} related to each of these receptors is specific to the chemical properties of the ligand. Here, we distinguish between the concentration of a receptor (e.g., $[ER\alpha 66]$), and the concentration of its activated form (e.g., $[ER\alpha 66^*]$). We assume receptor concentrations $[ER\alpha 66]$, $[ER\beta]$, and $[ER\alpha 46]$ are constant $([ER\alpha 66] = [ER\beta] = [ER\alpha 46] = 1$).

 $ER\alpha 66$ activity is modulated by ligand binding,

$$[\text{ER}\alpha 66^*] = \frac{V_{max}^{\alpha 66}[\text{ligand}][\text{ER}\alpha 66]}{[\text{ligand}] + K_m^{\alpha 66}},\tag{1}$$

where $V_{max}^{\alpha 66}$ and $K_m^{\alpha 66}$ are the typical Michaelis-Menten constants (gain and the half-maximum activation concentration, respectively). Other receptors share similar activation kinetics:

$$[\text{ER}\beta^*] = \frac{V_{max}^{\beta}[\text{ligand}][\text{ER}\beta]}{[\text{ligand}] + K_m^{\beta}},$$
(2)

$$[\text{ER}\alpha 46^*] = \frac{V_{max}^{\alpha 46}[\text{ligand}][\text{ER}\alpha 46]}{[\text{ligand}] + K_m^{\alpha 46}}, \text{ and}$$
(3)

$$[GPER^*] = \frac{V_{max}^{GPER}[ligand][GPER]}{[ligand] + K_m^{GPER}}.$$
(4)

The dynamics of the receptor activation, transcription, and proliferation are governed by a system of coupled ordinary differential equations. GPER concentrations are driven transcriptionally by $ER\alpha 66$ activation. This process is balanced by first-order turnover of the GPER protein,

$$\frac{d}{dt}[\text{GPER}] = k^{GPER} \left([\text{ER}\alpha 66^*] - [\text{GPER}] \right) \,. \tag{5}$$

Activation of ER α 36 is dynamic and is modulated by [GPER^{*}],

$$\frac{d}{dt}[\mathsf{ER}\alpha 36^*] = k^{\alpha 36} \left([\mathsf{GPER}^*] - [\mathsf{ER}\alpha 36^*] \right) \,. \tag{6}$$

Transcription has a base rate (zero-order) that is modulated by first-order turnover. Transcription rate is positively impacted by ER α 66, and negatively impacted by ER α 36 and ER α 46,

$$\frac{d}{dt}[\text{Transcription}] = k^{trans} \left(1 - [\text{Transcription}] + [\text{ER}\alpha 66^*] - [\text{ER}\alpha 46^*] - \phi_{\alpha 36}^{trans}[\text{ER}\alpha 36^*] \right) .$$
(7)

Proliferation (P) depends on transcription, $ER\alpha 36$, and $ER\beta$,

$$P = k^{P} \left([\text{ER}\alpha 36^{*}] + [\text{Transcription}] - [\text{ER}\beta^{*}] \right) \,. \tag{8}$$

The rate constants k^{GPER} , $k^{\alpha 36}$, k^{trans} , and k^P are set at 1 for simplicity. $\phi_{\alpha 36}^{trans}$ describes the relative contributions of ER $\alpha 36^*$ acting (a) through the kinase cascade (positive) and (b) through transcription (negative). To avoid degenerate coupling of these two mechanisms, it is important that $\phi_{\alpha 36}^{trans} \neq 1$. Here, we assume $\phi_{\alpha 36}^{trans} = 0.1$.

Equations 5–8 were evaluated in python using the scipy.integrate library with initial conditions as defined in Supplemental Table I. Five different ligands—each with distinct estrogen receptor binding behavior—were simulated to generate Figure 3B. ODEs were evaluated to their steady-state solutions (t = 2000), which were used Here, constant ligand concentrations were used and we have assumed no background level of receptor activation from endogenous estrogens. Neither of these assumptions are critical, and the model framework could be evaluated for more complicated inputs. For example, it may be appropriate to use a time-dependent (decaying) concentration of ligand for a system for which metabolism or clearance are known to be important. The effect of repeat dosing could be simulated by selecting an appropriate mathematical representation for [ligand]. Similarly, the impact of endogenous ligands on system dynamics could easily be considered.

When ligand concentration is constant (as is assumed in Fig. 3B), a steady-state solution to Eq. 8 can be determined analytically. The steady state value of GPER concentration, [GPER]^{ss}, follows from Eq. 5 as,

$$0 = k^{GPER} \left([\text{ER}\alpha 66^*] - [\text{GPER}]^{ss} \right)$$
$$[\text{GPER}]^{ss} = [\text{ER}\alpha 66^*].$$
(9)

The steady-state concentration of activated GPER, [GPER*]ss, follows from Eq. 4,

$$[GPER^*]^{ss} = \frac{V_{max}^{GPER}[\text{ligand}][ER\alpha 66^*]}{[\text{ligand}] + K_m^{GPER}}.$$
(10)

Similarly, from Eq. 6,

$$0 = k^{\alpha 36} \left([\text{GPER}^*]^{ss} - [\text{ER}\alpha 36^*]^{ss} \right)$$
$$[\text{ER}\alpha 36^*]^{ss} = [\text{GPER}^*]^{ss}$$
$$[\text{ER}\alpha 36^*]^{ss} = \frac{V_{max}^{GPER}[\text{ligand}][\text{ER}\alpha 66^*]}{[\text{ligand}] + K_m^{GPER}}.$$
(11)

And from Eq. 7,

$$0 = k^{trans} \left(1 - [\text{Transcription}]^{ss} + [\text{ER}\alpha 66^*] - [\text{ER}\alpha 46^*] - \phi_{\alpha 36}^{trans} [\text{ER}\alpha 36^*]^{ss} \right)$$

[Transcription]^{ss} = 1 + [ER\approx 66^*] - [ER\approx 46^*] - \phi_{\alpha36}^{trans} [ER\approx 36^*]^{ss} . (12)

Proliferation at steady state becomes,

$$P^{ss} = k^{P} \left(1 + (1 - \phi_{\alpha 36}^{trans}) [\text{ER}\alpha 36^{*}]^{ss} + [\text{ER}\alpha 66^{*}] - [\text{ER}\alpha 46^{*}] - [\text{ER}\beta^{*}] \right) \\ = k^{P} \left(1 + (1 - \phi_{\alpha 36}^{trans}) \frac{V_{max}^{GPER}[\text{ligand}] [\text{ER}\alpha 66^{*}]}{[\text{ligand}] + K_{m}^{GPER}} + [\text{ER}\alpha 66^{*}] - [\text{ER}\alpha 46^{*}] - [\text{ER}\beta^{*}] \right), \quad (13)$$

which depends on ligand concentration, but not time.

Variable	Initial State
[GPER]	1
$[ER\alpha 36]$	1
[Transcription]	1

Supplemental Table I: Initial conditions for simulation in Figure 3B.

	ER a 66		ERβ		ERa46		GPER	
	K_m	V_{max}	K_m	V_{max}	K_m	V_{max}	K_m	V_{max}
ER α 66-selective	1.0	1.0	1.0	0.2	1.0	0.1	0.5	0.5
nonspecific	1.0	1.0	0.1	0.3	1.0	0.1	0.5	0.5
ERβ-selective	1.0	0.1	0.1	0.3	1.0	0.1	0.5	0.5
$ER\alpha 46$ -selective	1.0	0.1	1.0	0.2	0.1	0.1	0.5	0.5
GPER-selective	1.0	1.0	1.0	0.2	1.0	0.1	0.01	0.5

Supplemental Table II: Parameter sets used for simulation in Figure 3B.