1 Ecological trait evolution in amphibian phylogenetic relationships

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- 22
- 23 Current biodiversity patterns of Neotropical amphibians are the result of their
- 24 functional and phylogenetic relationships. Understanding the associations between
- 25 ecological similarity and phylogenetic relatedness among species can provide a

26	convincing statement on the role of evolutionary history in the filling of the niche space.
27	Here, we assessed the ancestral character states of amphibian ecological traits and their
28	evolutionary history in the Atlantic Forest Hotspot. We used 12 genes (11,906 bp) to
29	reconstruct a phylogeny for 207 amphibian species and related it to eight ecological
30	traits regarding their morphology, life-history and behavioural features. We revealed
31	that closely related species can have similar ecological traits, suggesting that these traits
32	are driven by phylogenetic history. Despite the high endemism rate of Atlantic Forest
33	amphibians, our findings heavily rely on good studies on complete amphibian
34	phylogenetic lineages to overcome potential biogeographical constraints. Using
35	mechanisms of adaptive evolution in the context of phylogenetic diversification, we
36	suggest that closely related species have different phylogenetic signals and ecological
37	traits can evolve without relatedness.
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39	KEY WORDS: evolutionary changes, phylogenetic signal, functional niche, Atlantic
40	Forest, Anura, Gymnophiona.
41	
42	Running Head
43	Ecological trait evolution in amphibians
44	
45	INTRODUCTION
46	Ecological trait evolution is the main result of the evolutionary history of
47	functional interactions that addresses fundamental aspects on species' morphology, life-
48	history and behaviour, according to their phylogenetic relationships (Wiens & Graham
49	2005; McGill et al. 2006; Kraft et al. 2007; Violle et al. 2007). As closely related
50	species are often functionally similar (Losos 2008), many ecological traits show strong

phylogenetic signals (Freckleton et al. 2002; Webb et al. 2002; Moles et al. 2005; 51 52 Donoghue 2008). In this context, a central question is how similar do traits need to be to qualify them as conserved (Pyron et al. 2015). The magnitude of phylogenetic signals 53 can indicate the degree to which closely related species tend to have similar traits 54 (Blomberg et al. 2003). Empirical evidence suggests that within a major clade, some 55 lineages show strong phylogenetic signals in ecological traits, whereas others are likely 56 to show predominantly divergent traits (Diniz-Filho et al. 2010). A single ecological 57 trait can show different values of phylogenetic signal in relation to which node is 58 considered to evaluate it (Swenson & Enquist 2009). Therefore, systematic measures 59 60 that allow an evaluation how closely related species tend to retain more similar traits than distantly related may provide key insights into the evolutionary processes behind 61 ecological patterns (Duarte et al. 2012). 62 63 One of the main challenges in evolutionary biology is to explain the relationships of ecological traits in functional niches, and how these traits can change 64 multiple times (Gomez-Mestre et al. 2012). Another challenge which has yet to be 65 addressed is to understand how evolutionary traits are related with the long-term 66 maintenance of ancestral characters and the multiple origins of common ancestral states 67 (e.g. Hansen & Houle 2004; Uyeda et al. 2011). The key strategy to address these 68 69 challenges is assessing the relationships between ecological and phylogenetic traits (Cadotte et al. 2009; Gravel et al. 2011). Understanding the associations between 70 ecological similarity and phylogenetic relatedness is an essential step to reveal 71 72 biodiversity assembly mechanisms and ecosystem functioning (Hof et al. 2010; Bello et al. 2017). 73 Current patterns of diversity and distribution of tropical amphibians is a 74

consequence of their ecological and phylogenetic traits (Jetz & Pyron 2018).

76	Evolutionary processes involving amphibians at a macroscale have been widely
77	explored in previous studies (e.g. Wiens 2007; Hof et al. 2010; Ernst et al. 2012; Fritz &
78	Rahbek 2012; Duarte et al. 2014; Trakimas et al. 2016). Some studies also highlight a
79	tendency of amphibians to exhibit some degree of phylogenetic signal in their
80	ecological traits (Wiens et al. 2006; Wollenberg et al. 2008; Moen et al. 2009).
81	Incorporating phylogenetic information into trait-based approaches can be useful tools
82	for predicting ecological processes (Cavender-Bares & Wilczek 2003), such as
83	environmental filtering on amphibian community dynamics (Ernst et al. 2012).
84	Here, we test whether closely related species are more similar in their ecological
85	traits than expected if traits are independent of species phylogenetic relatedness, using
86	the amphibians of the Atlantic Forest Hotspot as a case study. We calculated
87	phylogenetic signals in ecological traits and provide evidence that these traits follow a
88	Brownian evolution model. In this context, we determine whether the ecological traits
89	of the Atlantic Forest amphibians are potentially driven by phylogenetic relationships.
90	Our findings reveal that phylogenetically related species may have different ecological
91	functions and the strength of the phylogenetic signals can vary across amphibian orders,
92	families and subfamilies.
93	
94	MATERIALS AND METHODS
95	Study area
96	The Atlantic Forest is one of the five most important biodiversity hotspots on
97	Earth (Myers et al. 2000). Originally, it covered around 1,500,000 km ² of which only
98	about 12% (~ 195 km ²) remains in Brazil, Paraguay and Argentina (Ribeiro et al. 2009),
99	corresponding to around 100,000 km ² of Brazilian forest remnants (Tabarelli et al.

100 2005). These forest remnants face a high rate of habitat loss (Teixeira et al. 2009),

101	which is one of the main factors driving amphibian populations to extinction (Stuart et
102	al. 2004; Becker et al. 2007; Ferreira et al. 2016). Despite this, the Atlantic Forest is still
103	considered the leading biome in terms of amphibian diversity in Brazil (Haddad et al.
104	2013), harbouring more than 50% of all amphibian species listed in this country
105	(Haddad et al. 2013). Its geographical characteristics including a wide altitudinal range
106	has favoured high species richness and levels of endemism, including more than 543
107	recognized amphibian species (Haddad et al. 2013). Here, we used the term Atlantic
108	Forest according to the vegetation remnant map produced by the SOS Mata
109	Atlântica/INPE (2015).
110	
111	Data acquisition
112	We obtained spatial data on amphibian species following two approaches. First,
113	we built a data set covering all currently recognized taxa known from the Atlantic
114	Forest following Haddad et al. (2013); and second, we performed complementary
115	fieldwork in seven protected areas across the Serra do Mar Coastal Forests and the
116	Central Corridor of the Atlantic Forest, stretching from the South to the Northeast of the
117	country (see Fig. S1) to compile additional data on species distribution and ecological
118	traits. In all surveyed localities, we used acoustic and visual nocturnal/diurnal
119	assessments (Crump & Scott 1994; Zimmerman 1994), through an active search for
120	amphibians around water bodies, streams and along 2,000 m forest transects within each
121	protected area. Following procedures approved by the American Society of
122	Ichthyologist and Herpetologist (ASIH 2004), we euthanized the collected species with
123	hydrochloride benzocaine (≥ 250 mg/L), under the national biological sampling between
124	2015-2018 (ICMBio-SISBIO; license #30344/44755).

126 Ecological traits

We characterized 207 amphibian species according to eight ecological traits that 127 determine different dimensions of the amphibians' functional niches in relation to their 128 morphology, life-history and behaviour. We used the following traits according to 129 130 Haddad et al. (2013), with some additional complements obtained in the fieldwork: (1) Activity (nocturnal; diurnal; nocturnal and diurnal); (2) Body size (small < 3 cm; 131 medium 3-10 cm; large > 10 cm); (3) Calling site (without calling; bamboo groove; 132 133 swamp and pond; bromeliad; forest floor; tree crown; cave and burrow; stream and 134 rivulet; river; low vegetation; leaf titter; swamp, pond, stream and rivulet); (4) Toxicity 135 (unknown; toxic; unpalatable; non-toxic); (5) Habit (arboreal; terrestrial; cryptic; fossorial; rheophilic; semi-aquatic; aquatic; arboreal and cryptic; arboreal and aquatic; 136 fossorial and aquatic; arboreal and terrestrial); (6) Habitat (forest; open area; forest and 137 open area); (7) Developmental mode (direct; indirect); (8) Members (apod; tetrapod). 138 139 Given the different modes of adaptive evolution at phylogenetic lineages both among "apods" and among "tetrapods", we used the category of "Members" in order to resolve 140 the evolutionary origins between the Anura and Gymnophiona species. Considering the 141 142 functional similarities among the ecological traits of the species evaluated, we used some non-exclusive character states, which can be represented by more than a single 143 character. Such ecological traits contribute to ecosystem supporting services through 144 direct and indirect changes on the ecosystem functions and processes (Hocking & 145 Babbitt 2014). These functions can be structural (habitat and habit) and ecological 146 (body size, members, activity, toxicity, calling site, and developmental mode). For 147 further details, see the Supporting Information (Table S1), where we showed the 148 149 specific functions and the ecosystem supporting services of each one of the ecological

traits assessed (Duellman & Trueb 1994; Toledo et al. 2007; Wells 2007; Haddad et al.

151 2013; Hocking & Babbitt 2014).

152

153 *Phylogenetic tree*

154 According to the time-calibrated phylogetic framework proposed by Jetz & Pyron (2018), we used an existing molecular supermatrix to produce a novel phylogeny 155 estimate for the Atlantic Forest amphibians (i.e. 11,906 bp for each species), through 156 157 three mitochondrial (i.e. Cyt-b, 12s and 16s) and nine nuclear genes (i.e. CXCR4, H3A, 158 NCX1, POMC, RAG1, ROHD, SIA, SLC8A3 and TYR). For the length-variable 159 regions, we performed multiple pairwise comparisons by the online version of MAFFT v.6.8 and the G-INS-i algorithm (Katoh & Toh 2008). After, we put together alignments 160 of all genes in the same alignment using the software SequenceMatrix 1.7.7 (Vaidya et 161 al. 2011) to concatenate the supermatrix previously produced. We obtained the 162 163 molecular data of 207 species in terms of nucleotide sequences obtained from GenBank (Benson et al. 2013; Table S2), provided by the National Center for Biotechnology 164 Information (NCBI). 165 166 We inferred the phylogenetic relationships performing Bayesian analyses in software BEAST 1.7 (Drummond et al. 2012), which was based on the combined data 167 matrix, a HKY model of sequence evolution for all genes, using a Yule speciation 168 process as the tree prior and an uncorrelated relaxed clock. We run the Yule process for 169 100 million generations, ensuring that the number of generations convergence were 170 sufficient assessed with Tracer 1.6 (Rambaut et al. 2014), removing a conservative 10% 171 burn-in fraction for the final tree. We combined these results with the use of 172 LogCombiner 1.8 (Rambaut & Drummond 2013a). We conducted additional 173 174 phylogenetic estimations based on 100 stochastic trees to account for the phylogenetic

175uncertainty of the single reconstructed phylogenetic tree for our 207 species assessed.176Given these 100 random simulations, we built the maximum clade credibility tree177(summary tree) with the use of TreeAnnotator 1.8 (Rambaut & Drummond 2013b). To178account node support, we used posterior probability of each node in accordance with the179100 trees generated automatically. We considered the nodes strongly supported if they180received posterior probability ≥ 0.95 .

The reconstructed phylogenetic tree provides a revised taxonomic classification 181 that incorporates the newly obtained phylogenetic information slightly changing in an 182 existing classification (Frost 2019). Thus, we did not take the low supported nodes as a 183 184 reason of changes in the current classification. We only used species from one geographic region, which in case of low node support (not shown), could be a result of 185 186 an incomplete lineage sorting and thus the lack of information in the sequence data. 187 We estimated the phylogenetic relationships of the ecological traits by using the most recent maximum-likelihood topology for 7,238 extant amphibian species, 188 according to the time-calibrated reconstruction proposed by Jetz and Pyron (2018). To 189 190 edit our phylogenetic tree based on this recent time-calibrated reconstruction, we used

191 the package 'ape' (Paradis 2004), in the R software (R Development Core Team 2017).

Finally, we revealed a wide-ranging phylogenetic tree across two orders, 17 families and15 subfamilies (Fig. 1).

194

195 Reconstruction of ancestral character states

We reconstructed the ancestral character states using maximum-likelihood
estimations under stochastic character mapping analysis (SIMMAP, Bollback 2006),
using 1,000 simulations for discrete characters based on the ecological-trait matrix
(Table S3). We compared the likelihood estimations with a value of lambda = 1 for

200	providing a standardized method to draw the reconstructed phylogenies on the
201	distribution of ecological traits, accounting the phylogenetic uncertainty for the
202	ancestral character states. We also used null simulations across the tips of the
203	reconstructed trees indicating the expected variances within the ancestral characters
204	under Brownian motion. We performed the stochastic character mapping analysis and
205	the null simulations in the R software (R Development Core Team 2017), using the
206	"make.simmap" and "fastBM" functions of the package "phytools" (Revell 2012).
207	Given that the characters in common for several lineages can originate from the
208	common ancestor of those lineages and may converge to the tips of the tree (Pavoine et
209	al. 2008), we assessed the relationships among the diversity of ancestral states at
210	different taxonomic levels. Resolving the evolutionary origins of the ecological traits,
211	we used an additive partitioning of diversity based on the ancestral states observed and
212	expected for the taxonomic levels "species", "subfamilies", "families", and "orders".
213	For each taxonomic level, a value of alpha diversity was calculated (i.e. the number of
214	character states). Therefore, $\alpha 1$ represented the alpha diversity in the lower taxonomic
215	level (species), so that $\alpha 2$, $\alpha 3$ and $\alpha 4$ corresponded to alpha diversity in the three
216	subsequent levels (subfamilies, families and orders). According to the protocol
217	proposed by Crist et al. (2003), we tested whether the distribution of diversity across
218	levels differs from the expected by chance using a null model under a 95% confidence
219	interval with 999 randomizations. We used expected diversity partitions through the
220	null model implemented in the "r2dtable" function, which is based on the Patefield
221	algorithm for generating random matrices of diversity across levels (see Blüthgen et al.
222	2008). We performed all analyses for the additive partitioning of diversity in the R
223	software (R Development Core Team 2017), using the algorithms "boot" and "mass"
224	through the "adipart" function of the package "vegan" (Oksanen et al. 2013).

225	We evaluated the ancestral traits for the following "lineages" from the original
226	tree: Orders – Gymnophiona and Anura; Families – Typhlonectidae, Siphonopidae,
227	Microhylidae, Leptodactylidae, Pipidae, Ranidae, Aromobatidae, Hylodidae,
228	Cycloramphidae, Bufonidae, Craugastoridae, Brachycephalidae, Eleutherodactylidae,
229	Odontophrynidae, Hemiphractidae, Phyllomedusidae and Hylidae; and Subfamilies -
230	Gastrophryninae, Paratelmatobiinae, Leiuperinae, Leptodactylinae, Allobatinae,
231	Craugastorinae, Holoadeninae, Ceuthomantinae, Phyzelaphryninae, Hemiphractinae,
232	Scinaxinae, Pseudinae, Lophyohylinae, Dendropsophinae and Cophomantinae. We
233	followed Frost (2019) for the taxonomic nomenclature of families, subfamilies and
234	species.
235	
236	Phylogenetic signal
237	We assessed the phylogenetic signal of each ecological trait using a robust test
238	proposed by Abouheif (1999). The Abouheif's C_{mean} test uses the Geary's C and
239	Moran's I indices, providing a phylogenetic proximity matrix that does not relate to
240	branch length but focuses on topology of the tree and has a non-zero diagonal values
241	(see Pavoine et al. 2008). We estimated Abouheif's C_{mean} with 999 randomizations
242	using the package "adephylo" (Jombart et al. 2010), in the R software (R Development
243	Core Team 2017).
244	In order to test what ecological traits follow a stochastic Brownian evolution
245	model, we used a measure based on the maximum likelihood of phylogenetic signals,
246	called lambda (λ), developed by Pagel (1999). This metric is a scaling parameter that
247	measures the phylogenetic dependence of observed trait data as a multiplier applied to
248	the internal branches of the tree. The lambda parameter ranges from 1 to 0, where $\boldsymbol{\lambda}$
249	equal to 1 correspond to the Brownian evolution model, whereas λ equal to 0

correspond to a completely stochastic distribution within the tree branches and an absence of phylogenetic signal (Münkemüller et al. 2012). The strongest argument for using Pagel's λ is that it provides a reliable effect size measure besides testing for phylogenetic signal for both continuous and discrete traits (e.g. Gumm & Mendelson 2011; Münkemüller et al. 2012; Best & Stachowicz 2013). For this analysis, we used the "phylosig" function of the package "phytools" (Revell 2012), in the R software (R Development Core Team 2017).

The main reason behind our choice by the Abouheif's C_{mean} and the Pagel's λ 257 tests is their high power to reduce uncertainty besides testing for phylogenetic signal in 258 259 large phylogenies, providing a level of performance substantially better than other methods (i.e. Fritz & Purvis' D or Blomberg's K; Münkemüller et al. 2012). Fritz & 260 261 Purvis' D test is a statistic approach to measure phylogenetic signal strength only in 262 binary traits, which are not the encountered in our case study. In this context, we followed the guidelines proposed by Münkemüller et al. (2012) to better assess 263 phylogenetic signal and distinguish it from random trait distributions. Given that 264 Pagel's λ is most valuable for discrete traits that follow Brownian motion (Best & 265 Stachowicz 2013), we fitted the reconstructed phylogenetic trees to a model selection 266 approach based on the delta Akaike Information Criterion (AAIC) weights for three 267 268 evolutionary models (i.e. BM = Brownian-motion model; EB = Early-burst model; White = White-noise model). We performed these models of character trait correlated 269 evolution using the "fitDiscrete" function of the "geiger" package (Harmon et al. 2007), 270 271 in the R software (R Development Core Team 2017). 272 In addition to the measures described above, we created a traitgram to visualize the evolutionary diversification for the ecological traits across the reconstructed 273

274 phylogeny under ancestral character estimation by likelihood, using the "phenogram"

275	function of the package "phytools" (Revell 2012), in the R software (R Development	
276	Core Team 2017). Time scales analysis of the multigene dataset are congruent with	
277	previous studies (San Mauro et al. 2005; Roelants et al. 2007; Wiens 2007; Blackburn et	
278	al. 2010; Roelants et al. 2011) and support anuran radiation episodes in the	
279	Triassic/Early Jurassic (basal anuran radiation).	
280		
281	RESULTS	
282	Maximum-likelihood ancestral state reconstructions suggested different	
283	evolutionary patterns on the distribution of ecological traits, fitting the Brownian-	
284	motion evolution model (Fig. 2). Activity trait indicated the "nocturnal" character as an	
285	ancestral state shared by the orders Anura and Gymnophiona, and the "diurnal"	
286	character as a derived state for the families Aromobatidae, Hylodidae and	
287	Brachycephalidae (exclusive for the genus Brachycephalus) (Fig. 2a). Body size	
288	indicated the "large (> 10 cm)" character as an ancestral state shared by the orders	
289	Anura and Gymnophiona, and the "medium (3-10 cm)" and "small (< 3 cm)" characters	
290	as derived states for the order Anura (Fig. 2b). Calling site and Toxicity showed derived	
291	states among anuran families due to emerged independently through different ancestors	
292	(Fig. 2c-d). Habit trait indicated the "aquatic" character as an ancestral state for the	
293	subfamily Pseudinae, and the "arboreal" character as a derived state for the families	
294	Hylidae, Hemiphractidae and Phyllomedusidae (Fig. 2e). Habitat trait indicated all	
295	characters as derived states ancestrally from forest environments (Fig. 2f).	
296	Developmental mode indicated the "direct" and "indirect" characters as ancestral states	
297	(Fig. 2g). Members indicated the "apodal" character as a derived state for the order	
298	Gymnophiona, and the "tetrapod" character as an ancestral state for the order Anura	
299	(Fig. 2h).	

300 Additive partitioning of diversity showed the greatest number of ancestral states 301 at the lower taxonomic level (i.e. species), indicating multiple evolutionary origins and adaptive outcomes (i.e. 97% of total observed ancestral states; Fig. 3). Comparing these 302 results with the expected diversity provided by the null model, only the levels "species" 303 304 and "families" showed diversity values significantly higher than random expectations (P < 0.05). However, the levels "subfamilies", "families" and "orders" did not show 305 relevant diversity values, together accounting for less than 3% of the total diversity of 306 307 ancestral states evaluated.

Overall, Abouheif's C_{mean} and Pagel's λ tests indicated that all ecological traits 308 309 showed significant phylogenetic signals against random expectations. Results of the model selection approach based on the delta Akaike Information Criterion (ΔAIC) 310 311 weights support the Brownian-motion as the best fitted model for our trait evolution 312 approach (Table 1). The second best fitted model was the Early-burst (EB), due to some characters' change tends to be concentrated toward the base of the tree. According to 313 Abouheif's Cmean, the traits that had the highest values were developmental mode (Cmean 314 = 0.919, P < 0.001) and members (C_{mean} = 0.872, P < 0.001). Abouheif's simulations 315 represented the distribution of the statistical Cmean calculated from each ecological trait 316 along the phylogeny evaluated (Fig. 4). On the other hand, Pagel's λ also indicated the 317 318 highest values of the maximum likelihood for the traits developmental mode and members ($\lambda > 0.999$, P < 0.001), totally supporting a Brownian evolution model. The 319 traits body size, toxicity and activity also had high λ values ($\lambda > 0.900$, P < 0.001), 320 321 showing close relations to the Brownian evolution model as well. However, the traits 322 habit and habitat showed moderate phylogenetic signals under this same model ($\lambda >$

323 0.600, P < 0.001).

324	The traitgram visualised the evolution of traits through time, using a projection	
325	for the evolutionary diversification of all ecological traits assessed under ancestral	
326	character estimation by likelihood (Fig. 5). In addition, this estimate accounts for the	
327	uncertainty about ancestral states along branches and at nodes. Potential evolutionary	
328	shifts over longer time scales was based on the basal anuran radiation in the Triassic and	
329	Early Jurassic periods (i.e. about 200 million years ago). This projection illustrated a	
330	horizontal dimension of evolutionary divergence time, showing a quantitative trait	
331	evolution for the multiple ancestral characters of the Atlantic Forest amphibians.	
332		
333	DISCUSSION	
334	Our results showed a strong tendency to retain ancestral ecological traits in	
335	Atlantic Forest amphibians. Given the partitioning diversity of reconstructed ancestral	
336	states at taxonomic levels, we suggest the additive partitioning of ancestral states as an	
337	additional approach for the stochastic character mapping, in an attempt to reduce the	
338	uncertainty behind the evolutionary origin of the ecological traits among taxonomic	
339	groups. Despite this uncertainty, phylogenetic characteristics of some species can be	
340	influenced or directly affected by other not phylogenetically related species. Under	
341	similar ecological pressures, some species are reinforced to exhibit the same ecological	
342	traits, due to the high phenotypic plasticity of amphibians (Relyea 2001; Urban et al.	
343	2014; Delia et al. 2019), which might in turn imply a lower functional adaptability of	
344	species to current and future climate change (Urban et al. 2014).	
345	We highlighted Developmental mode (i.e. direct and indirect) and Members (i.e.	
346	apod and tetrapod) as the most ancestral traits across the long sequence of changes in	
347	the basal amphibian radiation at the Atlantic Forest. In many cases, indirect	
348	development characters seem to represent terminal stages that are retained for tens of	

millions of years without proceeding to direct development (e.g. Phyllomedusidae and 349 350 Hylidae families). Although we have shown high levels of homoplasy, some lineages do not seem to have been hampered in their ecological diversification. Many specialization 351 modes can be evolving with variations in different ecological traits related to behaviour, 352 353 habitat and developmental modes (Roelants et al. 2011). The development of the same ecological traits using different evolutionary mechanisms cannot be distinguished by 354 our results. However, most of specialization modes behind the ecological traits assessed 355 can have been widespread with homoplasy across parallel evolutions (see Bossuyt & 356 Milinkovitch 2000). 357

358 Despite an overall and robust observed trend of detection of phylogenetic signals in ecological traits, we found large variation in ancestral character histories among 359 360 taxonomic groups. Both measures used to determine the strength of the phylogenetic 361 signals were highly significant, suggesting that they are following a Brownian evolution model. This variation in ancestral character histories was supported by the length of the 362 branches (a surrogate of time), corresponding to the characters' rhythm of evolution 363 expected under a random walk model (Gingerich 2009; Hunt 2012; Hunt & Rabosky 364 2014). However, low phylogenetic signals in ecological traits (i.e. C_{mean} < 0.6) also 365 showed exceptions from the random walk model, especially in the terminal branches. 366 367 These exceptions may indicate that ecological condition changes could be fast in these taxonomic bifurcation points and not in the overall reconstructed tree. On one hand, 368 mapping trait evolution on reconstructed phylogenies allows estimating where the 369 370 phylogenetic nodes connect the species trait values to the trait values of their potential 371 ancestors (Ackerly 2009; Kembel et al. 2010; Revell 2012). On the other hand, the 372 phylogenetic relationships of species showed by the traitgram can be difficult to 373 understand, mainly when species in different parts of the phylogeny have similar

ecological traits (Revell 2013). In this context, Campos et al. (2017) revealed a high
congruence of functional and phylogenetic patterns of amphibian biodiversity,
providing potential trade-offs for ecological and evolutionary processes in the Brazilian
Atlantic Forest.
Some other studies stated a doubt if the establishing of the existence of a

phylogenetic signal is a useful approach for integrating evolution, ecology, and
conservation biology (Wiens & Graham 2005; Wiens 2008). However, there is an
ongoing demand for further evidence to identify phylogenetic signals in different
communities (Losos 2008). The attempts to assess the existence of a phylogenetic signal
are very relevant to researches of biodiversity gradients and species distribution models
for climate change predictions (Hof et al. 2010), while the fact of its generality is still
under discussion.

386 We observed that many ecological traits are conserved enough to show strong phylogenetic signals when closely related species have similar traits, whereas 387 convergence has occurred when species from different lineages have similar ecological 388 characters. Conserved traits have been observed in different amphibian lineages, at least 389 within lower clades such as families or genera (Wiens et al. 2006; Wollemberg et al. 390 2008; Algar et al. 2009; Moen et al. 2009). Among these clades, the Hylidae family 391 392 showed to be generally conserved across the evolutionary history of the group in relation to climatic factors (Wiens et al. 2006). This association with speciation rates 393 can be explained by latitudinal diversity gradient (Wiens et al. 2011). Some 394 395 salamanders of the genera Plethodon and Desmognathus also showed relationships between speciation and endemism rates with conserved traits across the phylogeny 396 (Kozak & Wiens 2006). The widespread conservatism hypothesis is supported by 397 phylogenetic clusters based on ecological niche constraints (Pianka et al. 2017) and 398

their geographic location (Wiens et al. 2006). However, we need to take into account 399 400 that most associations between phylogenetic and ecological traits are scarcely based on an entire phylogenetic tree (Diniz-Filho et al. 2010). Therefore, some phylogenetically 401 conserved traits apparently attributed to regional amphibian clades can be 402 403 underestimated due to history-related biogeographical constraints on different phylogenetic lineages (Hof et al. 2010; Ernst et al. 2012). 404 We showed a straightforward and promising approach in how amphibian 405 ecological traits can recover significant phylogenetic signals in the Atlantic Forest. 406 However, our findings heavily rest on good studies on complete amphibian 407 408 phylogenetic lineages to overcome potential biogeographical constraints. Our study includes basically all Atlantic Forest amphibian species with available data on both 409 410 phylogenetic and ecological features, highlighting how badly more basic research is 411 needed to provide empirical data for testing evolutionary and ecological questions. Although we found that closely related species can show different ancestral 412 states, we revealed that most amphibians of the Atlantic Forest have ecological traits 413 414 driven by phylogenetic history. However, the strength of their phylogenetic signals varied considerably across amphibian orders, families and subfamilies. In summary, we 415 used a novel approach for investigating reconstructed ancestral states under maximum 416 417 likelihood and phylogenetic signals across an entire class of organisms in the Atlantic Forest, accounting differences from lower (species) to higher (orders) taxonomic levels. 418 Despite the potential biogeographical constraints of our assumptions, our results 419 420 address how the ecological trait evolution of amphibians can be informative to describe regional phylogenetic patterns based on multiple and discrete characters. A starting 421 422 point to address questions related to conservatism hypothesis and biogeographical constraints of different phylogenetic lineages may help to describe the evolutionary 423

424	patterns that may be important for environmental filtering. This work has sought to
425	move forward the use phylogenetic signals as a proxy for ecological similarities,
426	supporting conservation studies that explore ecological drivers of phylogenetic loss in
427	biodiversity hotspots.
428	
429	ACKNOWLEDGMENTS
430	We especially thank the Asociación Española de Ecología Terrestre (AEET) for
431	the research award granted to conduct this research. We are grateful to L. Rincón, P.
432	Pintanel, A. Beckerman, and two anonymous reviewers for the constructive comments
433	and great suggestions on the paper. We also thank the Institut de Biologia Evolutiva
434	(CSIC-UPF) for making the use of lab computers available. All data needed to evaluate
435	the conclusions in the paper are present in the paper and/or the Supporting Information
436	(Appendix I). Additional data related to this paper may be requested from the authors.
437	
438	DISCLOSURE STATEMENT
439	No potential conflict of interest was reported by the authors
440	
441	FUNDING
442	This research work was supported by the Coordination for the Improvement of Higher
443	Education Personnel (CAPES: 99999.001180/2013-04), and the National Council for
444	Scientific and Technological Development (CNPq: 140710/2013-2).
445	
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449	AUTHOR CONTRIBUTION	
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452	designed the analyses, collected the data, and created the figures. All co-authors	
453	discussed the results and edited the manuscript.	
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Table 1.

733	Phylogenetic signal of ecological traits according to the Abouheif's C_{mean} and the
734	Pagel's λ tests for the Atlantic Forest amphibians. Results of fitting the delta Akaike
735	Information Criterion (ΔAIC) weights are calculated for three evolutionary models by
736	using $\lambda = 1$: BM = Brownian-motion model; EB = Early-burst model; White = White-
737	noise model (i.e. no phylogenetic signal).

Ecological traits	Abouheif's C _{mean} *	Pagel's λ^*	BM (ΔAIC)	EB (ΔAIC)	White (ΔAIC)
Body size	0.624	0.973	0.000	1.931	107.113
Calling site	0.550	0.949	0.000	6.736	233.852
Toxicity	0.645	0.973	0.000	0.860	217.514
Habit	0.489	0.824	0.000	0.438	241.406
Habitat	0.300	0.644	18.857	0.000	32.375
Developmental mode	0.919	0.999	2.194	0.000	197.221
Members	0.872	0.999	2.041	0.000	32.529
Wiembers	0.872	0.999	2.041	0.000	

- ^{*}All *P*-values < 0.001.

747 Figure Captions

748

- 749 Fig. 1. Reconstructed phylogenetic tree for 207 amphibian species of the Atlantic
- 750 Forest. Numbers indicate nodes of Families, and circles indicate nodes of Subfamilies.
- 751 Gymnophiona: 1. Typhlonectidae; 2. Siphonopidae; Anura: 3. Pipidae; 4. Microhylidae
- 752 (Gastrophryninae); 5. Ranidae; 6. Aromobatidae (Allobatinae); 7. Hemiphractidae
- 753 (Hemiphractinae); 8. Craugastoridae (Holoadeninae, Craugastorinae, Ceuthomantinae);
- 9. Eleutherodactylidae (Phyzelaphryninae); 10. Brachycephalidae; 11. Cycloramphidae;
- 755 12. Hylodidae; 13. Phyllomedusidae; 14. Hylidae (Scinaxinae, Pseudinae,
- 756 Lophyohylinae, Dendropsophinae, Cophomantinae); 15. Leptodactylidae
- 757 (Paratelmatobiinae, Leiuperinae, Leptodactylinae); 16. Odontophrynidae; 17.
- 758 Bufonidae.
- 759
- 760 Fig. 2. Maximum-likelihood ancestral state reconstruction for the amphibian
- recological traits of the Atlantic Forest (N = 207 species). (a) Activity; (b) Body size; (c)
- 762 Calling site; (d) Toxicity; (e) Habit; (f) Habitat; (g) Developmental mode; (h) Members.
- 763 Reconstructed phylogenetic trees show discrete traits through stochastic character

764 mapping (SIMMAP), based on based on 1,000 simulations. Vertical bars across the tips

- 765 of the trees indicate the expected variances on ancestral states among species under the
- 766 Brownian-motion evolution model.
- 767
- 768 Fig. 3. Partitioning diversity of the reconstruction of ancestral states observed and
- respected at different taxonomic levels (species, subfamilies, families, and orders),
- according to the reconstructed phylogenetic tree for 207 amphibians of the Atlantic
- 771 Forest. Random expectations are based on a null model under a 95% confidence interval

vith 999 randomizations. Bars are organized from lower (species) to higher (orders)

- 773 taxonomic levels.
- 774
- 775 Fig. 4. Phylogenetic signal simulations according to the Abouheif's test calculated
- for each ecological trait across the reconstructed phylogeny of amphibians of the
- 777 Atlantic Forest (N = 207 species). Black diamonds indicate the position of the observed
- mean distributions in relation to the C_{mean} randomizations. All *P*-values < 0.001.
- 779
- 780 Fig. 5. Traitgram showing a projection for the evolutionary diversification of
- 781 ecological traits across the reconstructed phylogeny of amphibians of the Atlantic Forest
- (N = 207 species). Y-axis represents the ecological trait values under ancestral character
- restimation by likelihood (log10-transformed). X-axis represents the relative time
- (millions of years) and the length of the branches represents a surrogate of time.
- 785 Transparent blue stain represents 95% of confidence interval. All *P*-values < 0.001.

Commented [U1]: The Figure 5 was updated